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Title: New therapeutic approach with Extracellular vesicles from stem cells for intestinal cystitis/bladder pain syndrome

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26 **ABSTRACT**

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Interstitial cystitis/bladder pain syndrome (IC/BPS) is a debilitating chronic disorder characterized by suprapubic pain and urinary symptoms such as urgency, nocturia, and frequency. The prevalence of IC/BPS is increasing as diagnostic criteria become more comprehensive. Conventional pharmacotherapy against IC/BPS has shown suboptimal effects, and consequently, patients with end-stage IC/BPS are subjected to surgery. The novel treatment strategies should have two main functions, anti-inflammatory action and the regeneration of glycosaminoglycan and urothelium layers. Stem cell therapy has been shown to have dual functions. Mesenchymal stem cells (MSCs) are a promising therapeutic option for IC/BPS, but they come with several shortcomings, such as immune activation and tumorigenicity. MSC-derived extracellular vesicles (MSC-EVs) hold numerous therapeutic cargos and are thus a viable cell-free therapeutic option. In this review, we provide a brief overview of IC/BPS pathophysiology and limitations of the MSC-based therapies. Then we provide a detailed explanation and discussion of therapeutic applications of EVs in IC/BPS as well as the possible mechanisms. We believe our review will give an insight into the strengths and drawbacks of EV-mediated IC/BPS therapy and will provide a basis for further development.

INTRODUCTION

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Interstitial cystitis/bladder pain (IC/BPS) syndrome is a debilitating pain syndrome that presents with a wide range of symptoms including pelvic pain, urinary frequency, urgency, and cystoscopy findings, such as Hunner's lesion or glomerulation following hydrodistension. However, its diagnosis depends on the exclusion of other overlapping disorders. The prevalence of IC/BPS was long known to be higher in females (1). However, recent reports have demonstrated a higher prevalence in males (2, 3). Although no consensus exists on the actual pathophysiology of IC/BPS, numerous theories have emerged including mast cell infiltration, inflammation, glycosaminoglycan layer/urothelial dysfunction, and autoimmune dysregulation (4, 5).

Urothelium, a special form of epithelial tissue that lines the urinary tract walls, including the proximal urethra and urinary bladder, serves as a vital barrier against pathogens, toxins, and wastes (6, 7). The urothelium, which is made up of uroplakins (UPs) complexes, forms the urine-blood barrier and is supported by a thick pseudostratified transitional epithelium (multi-layered) and an asymmetric and fully differentiated superficial membrane (umbrella) (8-10). UPs are categorized into four subtypes: UP1a, UP1b, UP2, and UP3, which are implicated in the urothelium's permeability. These proteins coalesce together to form crystalline plaques on the bladder lumen's surface (10, 11). Proliferation or hyperplasia is a defensive reaction to any damage to the urothelium to rebuild the urine-blood barrier (12, 13). The main obstacles to bladder tissue regeneration are the scarcity of adequate tissue sources and senescence-associated primary cultures of bladder cells for several passages (14). For example, some trials using gastrointestinal tract-derived tissue for bladder regeneration showed inefficiency due to tumor formation, recurrent infection, metabolic abnormalities, and stone formation

73 (15). Through the present comprehensive review article, we want to report risk and benefit of
74 stem cell therapy for IC/BPS. In addition, we would introduce new approach with EVs from
75 stem cell in order to push the limit of the stem cell therapy.

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CONTENTS

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79 **Conventional treatment for IC/BPS**

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81 There are several drugs and surgical methods which are recommended for IC/BPS therapy by
82 global societies, such as the American Urological Association (AUA), the European Society
83 for the Study of IC (ESSIC), and the Society of Interstitial Cystitis of Japan (SICJ) (16).
84 Unfortunately, the current conventional drugs and alternative surgical interventions do not
85 guarantee complete recovery and are associated with harmful side effects (16, 17).

86 Because of its chronic nature and high prevalence, bladder dysfunction is an attractive target
87 for stem cell therapy. Numerous preclinical trials for bladder dysfunction, such as detrusor
88 underactivity, stress urinary incontinence, overactive bladder, and IC/BPS, have been
89 established, although clinical investigations in patients are still sparse (18).

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92 **Urgent need of new therapeutical strategy**

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94 Stem cell-based bladder dysfunction therapy includes several mechanisms, such as anti-
95 inflammation, anti-fibrosis, urothelium regeneration, anti-oxidant, anti-apoptosis, and
96 modulation of specific signaling pathways including Wnt and AKT/mTOR pathways (19).

97 Therefore, stem cell-based therapy has garnered attention as a robust alternative option.
98 Mesenchymal stem cells (MSCs) have a proven record of therapeutic efficacy in human
99 clinical trials and have been effective in a wide range of pre-clinical studies of tissue
100 regeneration in various immunologic and degenerative diseases (20-22). MSCs secrete
101 paracrine factors, which are the key mediators of MSC-associated therapeutic activities. Most,
102 if not all, of the MSCs' paracrine activities, are mediated by extracellular vesicles (EVs),
103 which are 50–1000 nm in diameter and secreted by all cell types (23, 24). EVs pass through
104 biological barriers such as the blood-brain barrier (25) and synovial membranes (26). It is
105 evidenced that EVs are carriers of the exogenous RNAs, such as siRNA (27), miRNA (28),
106 and modified miRNAs (29), which could be functional molecules in vitro and in vivo. In
107 addition, previous reports demonstrated the immunostimulatory or immunosuppressive
108 capacities of EVs based on their target and cellular source (30). The immunomodulatory
109 capacities of EVs could be beneficial for the treatment of inflammatory, autoimmune, and
110 hypersensitivity diseases (31). Small EVs (sEVs), of 50 to 200 nm in diameter, isolated from
111 MSC culture supernatants that are maintained under various culture conditions, have proved
112 to be therapeutically effective in various preclinical models (23, 32).

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115 **Pathophysiology of IC/BPS**

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117 The aetiology of IC/BPS is perplexing (33). Nevertheless, numerous hypotheses endeavor to
118 explain IC/BPS pathogenicity. Bladder epithelial damage, mast cell activation,
119 neuroinflammation, suppression of tight junction protein, afferent nerve plasticity, infection,
120 abnormal urothelial signaling, destruction of the superficial urothelial glycosaminoglycan

121 (GAG) layer, and psychological factors have been reported as the etiological factors that lead
122 to IC/BPS as illustrated in Figure 1 (16, 34, 35). A commonly acknowledged postulation of
123 IC/BPS pathophysiology proposes that the chronic inflammatory state is induced by early
124 damage or defect in the mucosal membrane of the bladder. Urothelium, a unique type of
125 epithelium, is composed of polysaccharides (chondroitin sulfate and hyaluronic acid) in its
126 outer layers and glycoproteins in the deeper layers (36). The injured urothelium is the main
127 culprit for the impaired barrier function, which allows urine solutes, such as potassium ions,
128 to seep into the suburothelium, leading to neuronal and muscle cells depolarization and
129 inflammation-related damage, urgency, and pain (37, 38).

130 The bladder pain is aggravating, especially, during the bladder filling process. Besides its
131 barrier function, urothelium is also implicated in sensory transduction through sensing
132 physiological and chemical signals in the bladder wall and releasing signaling molecules (39).
133 Urothelium-mediated signal transduction is not fully characterized; however, urothelial cells
134 can produce substances P, acetylcholine, and ATP, which are involved in the activation of the
135 bladder afferent neurons (40).

136 Chronic inflammation possibly plays a key role in IC/BPS pathogenesis. In bladder biopsies
137 of some patients with bladder pain, mast cells, leucocytes, and lymphocytes were found
138 infiltrating the bladder wall and suburothelial layers, along with increased vasculature and
139 thickening of the bladder wall. The clinical observations demonstrated subsequent chronic
140 pain in patients with frequent reports of urinary tract infections (UTIs). UTIs are among the
141 exacerbating factors of IC/BPS that commence at an early age and progress to IC/BPS in
142 adulthood (41). In IC/BPS patients, the proliferating mast cells in the bladder wall have been
143 linked to inflammation, allergic responses, and bladder hypersensitization (42, 43).

144 Of note, a link has been shown between the incidence of clinical IC/BPS and autoimmune

145 diseases (44). Autoantibodies, which are involved in the autoimmunity mechanism, showed
146 an adverse action on the bladder urothelium, connective tissues, and smooth muscles (44).
147 Further, a nationwide study recently reported IC/BPS in patients with primary Sjögren's
148 syndrome (45). Taken together, there is a strong link between autoimmune disorders and
149 IC/BPS pathogenicity, as evidenced by the chronic inflammation and the presence of
150 autoantibodies.

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152 **Limitations of stem cell therapy**

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154 Before discussing stem cell therapy limitations, we first need to briefly address the beneficial
155 effects of stem cells in the treatment of various diseases, especially IC/BPS. Basically, MSC-
156 based therapies are attributed to their intertwined roles including suppressing inflammation
157 by releasing cytokines, supporting healing by expressing growth factors, altering host
158 immune responses by secreting immunomodulatory factors, augmenting responses from
159 endogenous repair cells, and acting as mature functional cells such as bone cells (46). Stem
160 cell-mediated IC/BPS therapy is ascribed to various mechanisms, including their direct
161 differentiation into the main bladder cells, including urothelium and smooth muscles (SMCs),
162 their transplantation via several routes, and the activation of the vital signaling pathways that
163 are involved in bladder regeneration, such as mitogen activated protein kinases, AKT, Wnt-
164 GSK3 β / β -catenin, and mTOR signaling pathways (16, 47-50). The possible application of
165 stem cells in treatment of the bladder diseases at the preclinical level is also proved (18).
166 However, Clinicaltrials.gov currently shows no ongoing stem cell treatment clinical trials in
167 IC/BPS (18). On the other hand, the main concern over stem cell therapy is the lack of safety
168 proof. It is difficult to implant foreign living cells into a sophisticated structure like the

169 human body. Due to the vision loss in patients with macular degeneration, an age-related eye
170 condition, after the injection of autologous stem cells at a U.S. clinic (51), there has been a
171 growing concern over the safety of unproven stem cell therapies are used. The risk of post-
172 transplantation tumorigenicity is associated with the donor's age, growth modulation by the
173 recipient tissues (52), and the dysfunction of the patient immune system due to long-term
174 chemotherapy (53). Of note, long-term in vitro culture of MSCs could lead to unfavorable
175 consequences, such as chromosomal abnormalities, senescence, and genetic instabilities,
176 which negatively influence the engraftment (52, 54). Moreover, investigations in rodents and
177 dogs demonstrated that intravenously injected MSCs are trapped in the pulmonary capillaries
178 and large populations of MSCs are largely cleared, but some get through to the damaged
179 target tissue (55-58). In addition, stem cell engraftment led to immune reaction-associated
180 stress that resulted in unfavorable outcomes, such as cellular necrosis and differentiation
181 anomalies (59).

182 Specifically, stem cell therapy holds numerous limitations and challenges that hinder its
183 clinical application in bladder disease therapies, such as the controversies over the
184 transplantation route and the dose of the cells, the undefined mechanism of action of several
185 stem cell-mediated bladder disease therapies, and the in vivo IC/BPS models used for
186 verification of stem cell effects lack the reproducibility and needs further careful
187 authentications (16, 60).

188 Furthermore, when stem cells are introduced to target cells, the therapeutic effectiveness of
189 MSCs may not correspond with engraftment, differentiation, or cell fusion (61). Overall,
190 MSCs therapeutic actions are mediated via the paracrine effect, which is involved in tissue
191 repair, and not replacement-based therapy (62). In this prime, numerous studies demonstrated
192 that MSC-conditioned culture medium produced therapeutic effects similar to cell delivery in

193 rodent models of various disease models (63, 64), which has been supported by genomics
194 data showing that MSCs secrete a huge array of bioactive proteins (65, 66).

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197 **Overcoming the hurdles with stem cell-derived EVs**

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199 MSCs application in urologic regenerative medicine has been widely studied due to their
200 multilineage differentiation capacity (67, 68). However, the potential MSCs tissue
201 regeneration mechanism is by their paracrine action via the released soluble factors including
202 growth factors, cytokines, and chemokines rather than MSC differentiation and structural
203 interaction with host tissue (69, 70). The paracrine effect is dependent on the transfer of
204 proteins, bioactive lipids, and genetic material such as mRNA, miRNAs, and other non-
205 coding RNA. The paracrine impact of stem cells is also mediated via secreted EVs (71).
206 MSCs can release a wide spectrum of soluble factors such as secretomes into the culture
207 medium, which are less immunogenic and tumorigenic (72, 73). MSC paracrine activity
208 could be classified into various activities, such as anti-apoptotic, anti-inflammatory,
209 angiogenic, immunosuppressive, and immunomodulating impacts (74).

210 The specific compositions of MSC-derived EVs (MSC-EVs) differ according to tissue source
211 and in vitro cell stimulation approach (72); EVs isolation procedures include
212 ultracentrifugation, filtration, immunoaffinity, precipitation, size exclusion, and microfluidic
213 devices (75, 76). The basic criteria for EVs isolation and characterization are based on the
214 recommendations by Minimal information for studies of extracellular vesicles 2018
215 (MISEV2018) (77). However, the recent EVs isolation techniques are limited by lack of
216 reproducibility and variation in the quality of the EVs produced, and therefore need further

217 considerations (78). Broadly, EVs could be categorized as "exosomes" or "microvesicles"
218 (24). The term "exosome" commonly refers to a specific class of sEVs (sEVs) formed by the
219 endosomal system (79), which vary from the "ectosomes" (microvesicles and microparticles)
220 that emerge from the plasma membrane (80) or other similarly sized EVs with an undefined
221 biogenesis pathway (81). "Small" in the term sEVs indicates a population range of
222 approximately 50–200 nm in diameter. The production of sEVs is currently thought to be one
223 of the mediators of MSC therapeutic properties (23). In numerous in vitro functional assays
224 and relevant pre-clinical disease models, sEVs derived from in vitro MSC cells have been
225 reported to have therapeutic activities that imitate those of MSCs (82, 83). Furthermore,
226 MSC-EVs convey a large amount of verified therapeutic agents into target cells, such as
227 nucleic acids, proteins, miRNA, and lipids, to modulate numerous biological functions (84).
228 MSC-EV-associated tissue regeneration is majorly attributed to the MSC-EVs capacity to
229 enhance the proliferation process and suppression of the apoptotic changes (85, 86). The
230 prominent role of MSC-EVs in maintaining the immune hemostasis is attributed to the
231 modulation of immune cell fate and inhibition of uncontrolled inflammation (85, 87). One of
232 the key mechanisms of MSC-EVs in tissue regeneration is stimulation of the angiogenesis via
233 activation of various signaling pathways (88, 89). The MSC-EVs urinary bladder wall
234 diseases therapeutic activities are mediated via the bladder tissues regeneration and
235 suppression of inflammation, which helps to prevent disease development and recurrence
236 (90). Besides the possible effects of the purified EVs in the treatment of bladder diseases, we
237 will also explain examples of the roles of stem cell conditioned medium (CM) or co-culture
238 platforms in bladder disease therapy that represent the paracrine action of stem cells in which
239 EVs are potently involved. In 2014, Adamowicz et al. demonstrated the regenerative capacity
240 of MSC-derived conditioned media (MSC-CM) in the bladder wall when administered

241 intravesical in IC patients, which was attributed to its high content of cytokines, growth, and
242 trophic factors that possess immunomodulatory, anti-inflammatory, and angiogenic activities
243 (91). In 2018, Xie et al. demonstrated the potency of umbilical cord-derived mesenchymal
244 stem cells (UC-MSCs) to alleviate the inflammatory-associated changes, enhance the
245 proliferation, and block the apoptosis when co-cultured within human urothelial cells, SV-
246 HUC-1 cells that were pretreated with TNF- α (92). Interestingly, the application of siRNA
247 targeting EGF in the UC-MSCs abolished the anti-inflammatory activity of UC-MSCs when
248 co-cultured with TNF- α -exposed SV-HUC-1 cells. The authors showed that increased AKT
249 and mTOR, as well as significant decrease in the protein expression level of the
250 cleaved caspase-3, are involved in the therapeutic activity of UC-MSCs against the in vitro
251 interstitial cystitis model using TNF- α -treated SV-HUC-1 cells (92).

252 Hypoxia is implicated in the activation of the inflammatory events and the consequent
253 fibrosis of the bladder smooth muscles (93). In this regard, a study by Wiafe and colleagues
254 detected the in vitro upregulation of hypoxia-associated genes, TNF- α , IL-1 β , IL-6, HIF3 α ,
255 VEGF, TGF- β 1, and α SMA, in 3% oxygen tension-exposed bladder SMCs and then tested
256 the effects of direct and indirect co-culture with bone marrow-derived MSCs (BM-MSCs) in
257 reducing hypoxia-related changes (94). Direct co-culture is reliant on cell-to-cell interaction,
258 whereas indirect co-culture (based on trans-well system) is cell-to-cell interaction
259 independent. Interestingly, both co-culture methods led to a significant downregulation of
260 TGF- β 1 and IL-6, which are associated with fibrosis and pro-inflammation, respectively.
261 Moreover, the significant increase in the expression level of the potent anti-fibrotic cytokine,
262 IL-10, and the marked decrease in the expression level of α SMA, collagen I and III
263 transcripts, and the total collagen proteins were demonstrated upon co-culture with BM-
264 MSCs. Taken together, MSC-EVs were evidenced to be effective IC therapies via several

265 mechanisms, including delivery of therapeutic miRNA and growth factors, anti-inflammation,
266 anti-fibrosis, and modulation of key signaling pathways (**Figure 2**).

267 In 2017, Lv et al. affirmed the cross-link between miR-214 inhibition and the enhancement of
268 epithelial-mesenchymal transition that leads to fibrosis of the bladder wall and subsequent
269 interstitial cystitis in postmenopausal women, which is mediated via the upregulation of
270 Mitofusin 2 (Mfn2) (95). Accordingly, another study confirmed the protective capacity of
271 exosomes derived from miR-214-enriched BM-MSC, which were cultured under hypoxic
272 conditions against the oxidative damage in the cardiac stem cells (96). This effect is mediated
273 via the suppression of calcium/calmodulin-dependent protein kinase II (CaMKII). Taken
274 together, this study paves the way for revisiting the effect of miR-214 as an attractive
275 candidate in exosome-mediated IC/BPS therapy. Various MSC-EVs showed anti-
276 inflammatory (97), anti-fibrotic (98, 99), immunomodulatory (100) functions via modulation
277 of a wide range of miRNAs, which need to be tested in IC/BPS therapy. **In addition, there is a
278 scarcity of studies that show the impact of stem cell derived EVs in IC/BPS therapy, which
279 paves the way for further studies and clinical trials as well.** A brief comparison between MSC
280 and EVs in cystitis therapy is summarized in Table 1.

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CONCLUSIONS

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286 IC/BPS is a complicated chronic illness with unclear etiology. There are a variety of IC/BPS
287 therapeutic options and surgical interventions, however, they are associated with detrimental
288 side effects and do not guarantee a complete recovery. MSCs have shown unique therapeutic

289 activities and have been considered for the treatment of bladder diseases. However, stem cells
290 engraftments for bladder tissue regeneration face major challenges such as immune reaction,
291 low survival rate, and tumorigenicity, which limit their clinical application. To overcome
292 these constraints, numerous scientific works have demonstrated the efficiency of paracrine
293 mechanisms of MSCs in the treatment of a wide range of diseases, which is represented in the
294 secretion of a diverse range of growth factors, miRNAs, proteins, cytokines, and chemokines.
295 EV secretion is considered the main mediator of MSCs paracrine mechanism. Here, we
296 highlighted the advantages of the application of EVs as a cell-free platform over the direct
297 use of stem cells in cystitis therapy. We explained how EVs contribute to cystitis treatment
298 via the enhanced proliferation, anti-inflammatory, anti-fibrotic, and immunomodulatory
299 functions. Furthermore, we showed the role of EVs as cargos for therapeutic molecules such
300 as miRNAs and their role in the alleviation of cystitis. However, further investigations into
301 miRNA carried by EVs in IC/BPS therapy are needed. Moreover, further in-depth
302 comparative studies on MSCs and their EVs in IC/BPS treatment are required for the
303 clarification of the effectiveness and efficacy of EVs over the parent MSCs before the clinical
304 applications. We believe further improvements in separation, characterization, engineering,
305 and efficacy evaluation of MSC-EVs are essential for deriving high quality EVs with utmost
306 activity for IC/BPS therapy in the future.

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316 **CONFLICTS OF INTEREST**

317 The authors declare no conflict of interest.

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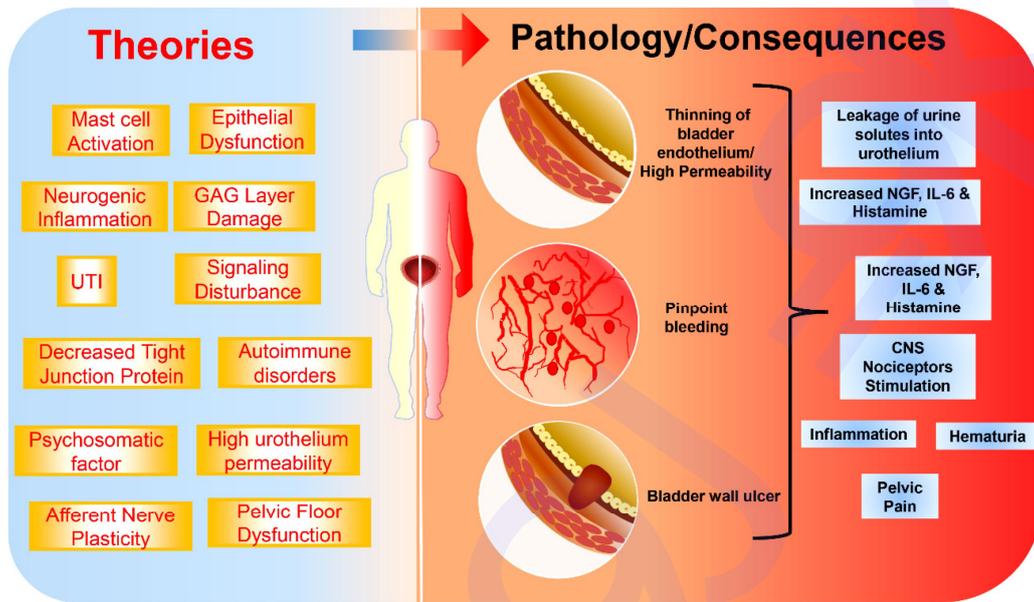
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FIGURE LEGENDS

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339 **Fig. 1. IC/BPS Pathophysiology**



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341 GAG: glycosaminoglycan, UTI: urinary tract infections, NGF: nerve growth factor, IL-6:
342 interleukin-6, CNS: Central nervous system

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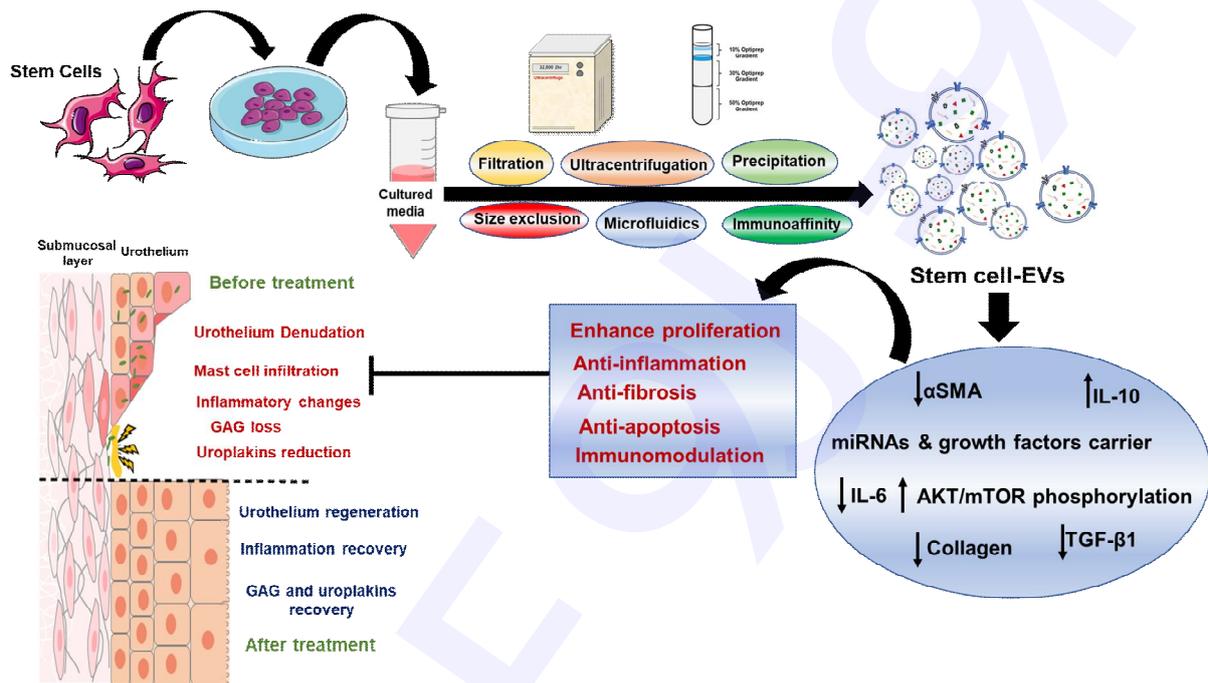
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349 **Fig. 2. MSC-EVs mechanisms in IC/BPS therapy.** A schematic diagram summarizing
 350 MSC-EV separation methods and their potential modes of action in IC/BPS therapy. Parts of
 351 this figure were created using Servier Medical Art (<https://smart.servier.com>), licensed under
 352 a Creative Commons Attribution 3.0.

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365 **Table 1.** Comparison of stem cell therapy and EVs therapy.

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Item	Stem cell therapy	EVs therapy
Source	Primary tissues of various body organs	Cultured media
Interstitial cystitis therapy mechanisms	<ul style="list-style-type: none"> -Cell replacement via their differentiation into urothelium and muscle layers -Migration to injured site -Anti-inflammation -Anti-fibrosis -Modulation of signaling pathways 	<ul style="list-style-type: none"> -Delivery of therapeutic molecules (miRNA and growth factors). -Enhance proliferation -Anti-fibrosis -Anti-apoptosis -Immunomodulation
Pros	<ul style="list-style-type: none"> -Availability -Easily isolated and expanded -Multilineage differentiation -Unique immunological properties 	<ul style="list-style-type: none"> -Cell-free platform -Carrier of the cell therapeutic molecules -Easily engineered. -Easily stored
Cons	<ul style="list-style-type: none"> -Post-transplantation tumorigenicity -Donor-dependent quality -Genetic instability and chromosomal abnormalities -Senescence -Short-term survival at the injured site -Engraftment failure 	<ul style="list-style-type: none"> -Costly -Irreproducible and inefficient separation methods -Heterogenicity -EVs characterization Difficulties -Scarcity of EVs specific markers

367 AKT: serine/threonine kinase, mTOR: mechanistic target of rapamycin kinase, TGF-β:
 368 transforming growth factor- β

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Fig. 1.

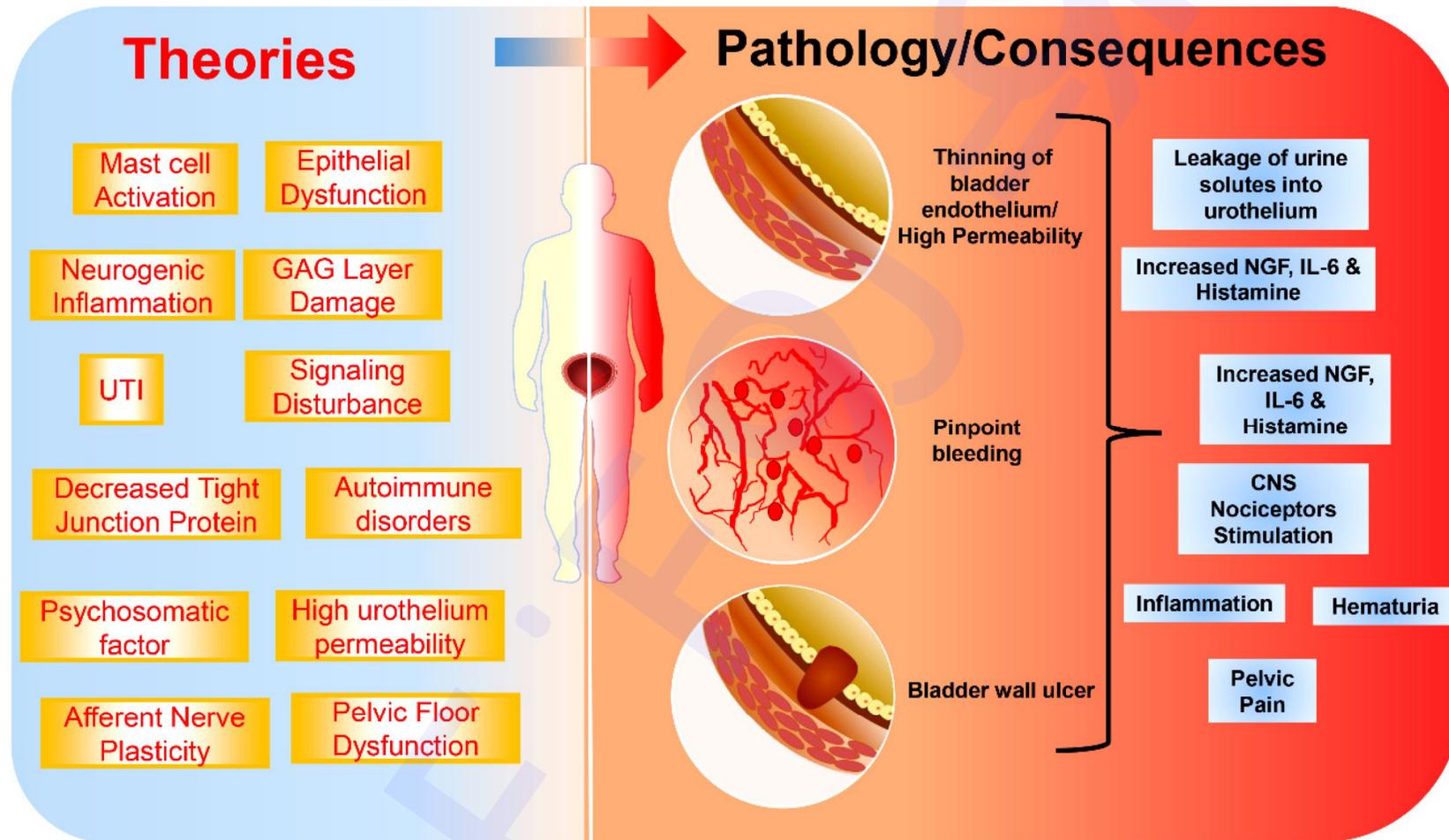


Fig. 2.

Stem Cells

