

BMB Reports – Manuscript Submission

Manuscript Draft

**Manuscript Number:** BMB-18-153

**Title:** Potential involvement of *Drosophila* flightless-1 in carbohydrate metabolism

**Article Type:** Article

**Keywords:** Flightless-1; gene expression; metabolomics; RNA sequencing; metabolism

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## Potential involvement of *Drosophila* flightless-1 in carbohydrate metabolism

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**Running title:** role of *Drosophila* flightless-1 in gene expression

**Keywords:** Flightless-1, gene expression, metabolomics, RNA sequencing, metabolism

**Abstract**

A previous study of ours indicated that *Drosophila* flightless-1 controls lipid metabolism, and that there is an accumulation of triglycerides in *flightless-1* (*fliI*)-mutant flies, where this mutation triggers metabolic stress and an obesity phenotype. Here, with the aim of characterizing the function of FliI in metabolism, we analyzed the levels of gene expression and metabolites in *fliI*-mutant flies. The levels of enzymes related to glycolysis, lipogenesis, and the pentose phosphate pathway increased in *fliI* mutants; this result is consistent with the levels of metabolites corresponding to a metabolic pathway. Moreover, high-throughput RNA sequencing revealed that *Drosophila* FliI regulates the expression of genes related to biological processes such as chromosome organization, carbohydrate metabolism, and immune reactions. These results showed that *Drosophila* FliI regulates the expression of metabolic genes, and that dysregulation of the transcription controlled by FliI gives rise to metabolic stress and problems in the development and physiology of *Drosophila*.

## Introduction

Flightless-1 was originally identified in *Drosophila* (1) and has been described as an actin-remodeling protein that belongs to the gelsolin protein superfamily (2-4). Flightless-1 contains a gelsolin-like actin-binding domain at the C terminus whereas a leucine-rich repeat (LRR) domain is located at the N terminus (1). The LRR domain is involved in both intramolecular recognition and structural organization. The gelsolin like domain mediates actin-binding and protein–protein interactions. The Flightless protein has been implicated in actin filament organization during cell migration and tissue repair (5). Beyond cytoskeletal function, flightless-1 has been shown to act as a transcriptional coregulator that can either positively or negatively affect the activity of transcription factors (6-9). Flightless-1 interacts with nuclear receptor (NR), transcription coactivators, and the SWI/SNF chromatin-remodeling complex. Flightless-1 binds to BAF53, a component of SWI/SNF complexes, as well as to estrogen receptor  $\alpha$  (ER $\alpha$ ), thus contributing to the recruitment of SWI/SNF complexes to the promoter of ER $\alpha$  targets (10). In addition, flightless-1 forms a complex with NR coactivators—glucocorticoid receptor–interacting protein 1 (GRIP1) and coactivator-associated arginine methyltransferase 1 (CARM1)—which leads to the enhancement of NR function (6). In contrast, flightless-1 inhibits  $\beta$ -catenin–mediated transcription through interfering with the binding of FLII leucine-rich repeat–associated protein 1 (FLAP1) to p300 and  $\beta$ -catenin (8).

A recent study of ours has shown that *Drosophila* flightless-1 (FliI) plays a role in lipid metabolism (11). *Drosophila fliI* mutants show increased levels of triglycerides and are resistant to starvation. In that study, the upregulation of FliI suppressed the mRNA expression of Desaturase-1, and conversely, the mRNA expression of Desaturase-1 increased in *fliI*-mutant flies, suggesting that *Drosophila* FliI downregulates Desaturase-1 at the transcriptional level, thereby contributing to the obese phenotype of *fliI* mutants. Based on

these results, our purpose here was to investigate the genes whose expression levels are regulated by *flightless-1* in *Drosophila*, thereby contributing to the above phenotype. Specifically, we examined the expression levels of genes related to energy metabolism, lipogenesis, and lipolysis; these genes may contribute to the obese phenotype, specifically, the increased levels of triglycerides and insulin resistance, in *fliI*-mutant flies. The mRNA expression of most glycolytic-enzyme genes and of some lipogenic enzymes was found to be specifically higher in *fliI*-mutant flies. Accordingly, we found that the levels of stearoyl-coenzyme A (CoA) and palmitoyl-CoA specifically increased in *fliI* mutants according to metabolomic analysis. In addition, high-throughput RNA sequencing (RNAseq) analysis of *fliI*-mutant flies revealed that *flightless-1* regulates the expression of genes related to chromatin organization, carbohydrate metabolic process, and proteolysis. Taken together, these results support the transcriptional role of *flightless-1* in the expression of genes associated with metabolism.

## Results

### Metabolic reprogramming in *fliI* mutants

Our recent study indicates that *Drosophila flightless-1* mutant flies contain large amounts of triglycerides, which contribute to starvation resistance (11). In general, fat accumulation has been primarily attributed to food intake and energy expenditure (12, 13). Based on the fact that our previous results revealed that *fliI* mutant flies do not consume more food (11), we tested whether or not FliI is required for the gene expression related to energy metabolism. To that end, we compared the transcript levels of glycolytic genes between seven-day-old controls and *fliI*<sup>3/14</sup> mutants. These *fliI* mutant alleles, *fliI*<sup>3</sup> and *fliI*<sup>14</sup>, have been characterized in previous studies (11, 14, 15). The *fliI*<sup>3</sup> mutant allele has a single-base substitution of Gly to Ser at amino acid position 602, which is homozygotically viable. In contrast, the *fliI*<sup>14</sup> allele is

lethal during the larval and pupal stages. The mRNA expression of most glycolytic-enzyme genes, including *HexA*, *Pgi*, *Gapdh*, *Pgk*, and *Eno*, specifically increased in *fliI*<sup>3/14</sup> mutants. Among them, the *Ald* transcript was upregulated more than sixfold in *fliI*<sup>3/14</sup> mutant flies, while the mRNA expression of *Ldh* significantly decreased (Fig. 1A). In contrast, the mRNA expression of tricarboxylic acid (TCA) cycle genes, *CG7430* and *Scsa*, did not significantly increase (Fig. 1B). These results showed that the mutation of *fliI* in *Drosophila* accelerated glucose metabolism throughout the body. 6-Phosphogluconate dehydrogenase (Pgd) in the pentose phosphate pathway, which is essential for the supply of NADPH (16), was also upregulated in *fliI*<sup>3/14</sup> mutant flies (Fig. 1C). The byproduct of NADPH produced by 6-Phosphogluconate dehydrogenase activity may be utilized for lipid synthesis. In addition, we found that the mRNA expression of lipogenic genes, such as sterol-regulatory element-binding protein (*SREBP*) and its downstream target genes, *fatty acid synthase (FAS)* and *acetyl-CoA carboxylase (ACC)*, was slightly higher in *fliI*<sup>3/14</sup> mutants (Fig. 1D). In line with these data, we found that the mRNA expression of DGAT1, which catalyzes the conversion of diacylglycerol and fatty acyl CoA to triglycerides, was significantly increased (Fig. 1D). In contrast, the mRNA expression levels of lipolytic genes, *brummer (Bmm)* and *hormone-sensitive lipase (HSL)*, did not significantly change (Fig. 1E). Altogether, these results suggest that *FliI* is necessary for the regulation of glucose metabolism and the transcription of lipogenic genes.

### **Metabolomic analysis of *fliI* mutants reveals a change in precursors of long-chain fatty acids**

In order to understand how changes in gene expression influence fat metabolism in *fliI* mutants, we compared the metabolic profiles of control and *fliI*<sup>3/14</sup> flies through mass spectrometry.

Specifically, we examined the intermediates of glycolysis, of the TCA cycle, of the pentose phosphate pathway, and the coenzymes involved in the metabolism of fatty acids. In agreement with the results of mRNA expression analysis by quantitative PCR, two important metabolic pathways, glycolysis and the TCA cycle, differed between *w<sup>1118</sup>* flies (controls) and *fliI<sup>3/14</sup>* mutants (Fig. 2A, B). Although lactate was shown to be downregulated in *fliI<sup>3/14</sup>* mutants, the amounts of glycolysis intermediates, such as fructose-1,6-bisphosphate, 3-phosphoglycerate, and phosphoenolpyruvate, increased relative to controls. *FliI<sup>3/14</sup>* mutants also manifested slight but significant increases in the amounts of TCA cycle intermediates, namely citrate/isocitrate, succinate, and fumarate. Nevertheless, the total ATP level did not differ between the two groups. In addition, we found that a pentose phosphate pathway intermediate, 6-phosphogluconate, was significantly upregulated in *fliI<sup>3/14</sup>* mutant flies. The amount of ribulose-1,5-bisphosphate, which is formed from ribulose 5-phosphate in the cooperative pentose phosphate pathway, significantly increased in *fliI<sup>3/14</sup>* mutant flies. A previous study suggests that ribulose-1,5-bisphosphate stimulates phosphofructokinase-1 and inhibits its opposing enzyme, fructose 1,6-bisphosphatase (17); this mechanism may contribute to glycolysis and the pentose phosphate pathway. NADPH is generated by the pentose phosphate pathway and is utilized for fatty acid synthesis. *FliI<sup>3/14</sup>* mutants also manifested a significant reduction in the ratio of NADPH/NADP (Fig. 2C).

The accumulation of long-chain acyl CoAs is frequently observed in obesity or type 2 diabetes (18, 19). Given that *fliI<sup>3/14</sup>* mutants had an obesity-like phenotype, we tested whether this phenotype in *fliI<sup>3/14</sup>* mutants directly affects the amounts of long-chain acyl CoAs. Although seven-day-old *fliI<sup>3/14</sup>* mutants showed decreased levels of coenzyme A and short-chain acyl CoA compared to the wild type, long-chain acyl CoAs, such as palmitoyl-CoA, oleoyl-CoA, and stearoyl-CoA, were found to be upregulated in *fliI<sup>3/14</sup>* mutant flies (Fig. 2D).

Differences in the expression levels of metabolic genes and relevant metabolites between  $w^{1118}$  flies (control) and  $fliI^{3/14}$  mutants, as well as the changes in  $fliI^{3/14}$  mutants are summarized in Fig. 3. In particular, increased levels of mRNA or a metabolite in  $fliI^{3/14}$  mutants are labeled in red, downregulation is indicated with blue, and unchanged ones are shown in black.

Given that *Drosophila* flightless-1 represses the expression of Desaturase-1 (11), whose preferred substrates are palmitoyl-CoA and stearoyl-CoA, we believe that the elevated amounts of long-chain acyl CoAs contributed to the obesity-like phenotype of  $fliI^{3/14}$  mutant flies.

### ***Drosophila* flightless-1 regulates the expression of genes related to chromatin remodeling**

In order to gain a genome-wide view of the changes in gene expression specifically induced by flightless-1, we performed a differentially expressed gene (DEG) analysis of RNA-seq data from seven-day-old  $w^{1118}$  control and  $fliI^{3/14}$  adult flies. Only the transcripts with adjusted  $P$  values (Benjamini-Hochberg correction)  $< 0.05$  and a fold change  $> 2$  in gene expression between the two groups were selected for subsequent Gene Ontology (GO) enrichment analysis. According to this criterion, 181 genes were downregulated in  $fliI$  mutants, while 160 genes were upregulated (see Table S1). The Gene Ontology (GO) terms with adjusted  $P < 0.05$  are listed in Tables 1 and 2. Specifically, GO terms enriched in the downregulated genes included those involved in chromosome organization (GO:0051276, adjusted  $P$  value of  $7.44 \times 10^{-5}$ ), the negative regulation of histone modification (GO:0031057, adjusted  $P$  value of  $8.39 \times 10^{-5}$ ), nucleosome assembly (GO:0006334, adjusted  $P$  value of  $9.87 \times 10^{-5}$ ), and a carbohydrate metabolic process (GO:0005975, adjusted  $P$  value of  $1.26 \times 10^{-4}$ ). This indicates that *Drosophila* FliI not only regulates specific gene expression related to glucose



metabolism, but also seems to regulate the gene expression in general under certain conditions by reorganizing chromatin structure. Prominent among the genes whose expression increased significantly were those encoding proteolytic enzymes (GO:0006508, adjusted P value of  $7.29 \times 10^{-7}$ ), proteins related to a defense response to a gram-positive bacterium (GO:0050830, adjusted P value of  $1.49 \times 10^{-3}$ ), and proteins associated with cytolysis (GO:0019835, adjusted P value of  $6.45 \times 10^{-3}$ ). The activators of Imd signaling (PGRP-LC) and lysozymes (LysB, -D, and -E) were highly expressed in *fliI* mutant flies (Table S1). An abnormally active immune response is associated with the upregulation of stress response factors, all of which have been previously linked to immune function, including the translational repressor 4E-BP/Thor (20). These data are in agreement with those of our recent study suggesting that the mRNA level of 4E-BP is increased in *fliI*<sup>3/14</sup> mutants (11). The RNA-seq. results were further validated through quantitative real-time PCR (supplemental Fig. 2). These observations indicate that the transcriptional activity of flightless-1 controls the expression of genes related to chromatin remodeling, metabolism, and immune responses.

## Discussion

Here, we reveal the transcriptional influence of *Drosophila* FliI on the expression of genes including those participating in chromatin remodeling, carbohydrate metabolic processes, and immune responses. Unexpectedly, our RNAseq analysis suggests that *Drosophila* flightless-1 regulates the expression of histones, which are basic nuclear proteins that are responsible for the nucleosome structure of the chromosomal fiber (21, 22). Histones are required for the condensation of nucleosome chains into higher-order structures. They also function as regulators of individual gene transcription through chromatin remodeling. In addition, the genes whose expression decreased in *flightless-1* mutants, *rhino* and *CREG*, are known to

control gene transcription through producing Piwi-interacting RNA (23) or inhibiting transcription factor binding (24). Thus, although *Drosophila* flightless-1, as a transcription factor, may regulate the expression of genes related to specific pathways including glucose metabolism, immune responses, or proteolysis, it seems to regulate gene expression by reorganizing the chromatin structure under certain conditions. These phenomena need to be validated through further studies.

Given that *fliI*<sup>3/14</sup> mutants show features of a metabolic disorder (11), we analyzed the changes in metabolites in the metabolic pathways including glycolysis, the TCA cycle, pentose phosphate pathway, and fatty acid synthesis (Fig. 2). Through this assay, we found that the NADPH/NADP ratio was clearly lower in the mutants. NADPH is an essential cofactor in *de novo* lipogenesis, which requires NADPH as a reducing agent for the conversion of acetyl-CoA into fatty acids (25-27). The amounts of NADPH in *fliI*<sup>3/14</sup> mutants were relatively low as compared to those in *w*<sup>1118</sup> flies. We can explain this result as a form of supply and demand: NADPH is mainly generated by the pentose phosphate pathway. Our results indicate that the level of ribulose 1,5-phosphate, which is a product of the pentose phosphate pathway, significantly increased in *fliI*<sup>3/14</sup> mutant flies (Fig. 2). In addition, mRNA expression of the gene encoding one of the NADPH-producing enzymes, phosphogluconate dehydrogenase (Pgd), increased slightly but significantly in *fliI* mutants (Fig. 1C). Therefore, we can theorize that in *fliI*<sup>3/14</sup> mutants, NADPH and acetyl-CoA were rapidly consumed for the elongation of long-chain fatty acids, driving the increase in fat contents throughout the body. Among the intermediates of glycolysis, the amount of lactate significantly decreased in *fliI*<sup>3/14</sup> mutant flies; this is consistent with the observed reduction in *Ldh* mRNA expression. As shown in Fig. 3, the levels of expression of glycolytic genes and glycolysis intermediates were significantly increased in *fliI*<sup>3/14</sup> mutant flies compared to in control flies. However, the mRNA expression levels of tricarboxylic acid (TCA) cycle genes and the levels of TCA

cycle intermediates were only slightly changed. Thus, we believe that the increased amounts of glycolytic intermediates were not consumed for lactate production but were instead mostly converted to acetyl-CoA for the elongation of long-chain fatty acids.

Given that *fliI*<sup>3/14</sup> mutant flies showed increased levels of triglycerides, we expected that the amounts of lipogenic enzymes, such as SREBP, FAS, and ACC, would be greater in *fliI*<sup>3/14</sup> mutant flies, but there were slight and significant differences in the expression of these proteins between *w*<sup>1118</sup> and *fliI*<sup>3/14</sup> flies. Emerging evidence indicates that the activity of a metabolic enzyme can be regulated post-translationally under certain conditions without changing the expression level (28-30). We assumed that an enzyme's activity can somewhat contradict its expression level because enzymes may be regulated by post-translational modifications in the mutant. Therefore, in future studies, it is necessary to test the activities of the enzymes that we examined in this study.

In summary, we analyzed the expression levels of metabolic genes and the levels of metabolites in *fliI*-mutant flies. These results support previous findings showing that *Drosophila* FliI serves as a key regulator of lipid metabolism. In addition, RNAseq analysis revealed the transcriptional targets of FliI in *Drosophila*, which include the genes related to chromosome organization, carbohydrate metabolism, proteolysis, and immune responses.

## Materials and methods

### Plasmids and fly strains

All *Drosophila* stocks were raised at 25 °C on a standard cornmeal medium containing 4.94% molasses, 3.8% cornmeal, 1.6% yeast, and 1.2% agar. Genes were expressed in *Drosophila* through the standard Gal4/UAS system. Fly strains *w*<sup>1118</sup> (stock number 5905), *fliI*<sup>3</sup> (stock

number 4730), and *fliI<sup>14</sup>/FM6* (stock number 7481) were obtained from the Bloomington Stock Center.

### Quantitative RT-PCR

Total RNA was isolated from five female flies using the TRIzol Reagent (Invitrogen, USA), and 200 ng of RNA was transcribed using the ReverTra Ace qPCR RT Kit (Toyobo Co., Japan). Quantitative PCR amplification was run for 40 cycles by means of the TOPreal™ qPCR 2X PreMIX (SYBR Green with high ROX) and a LightCycler® 480 Real-Time PCR System. *Rp49* served as a reference for normalization. Relative quantification of mRNA was performed through the comparative C<sub>T</sub> method. The primers are listed in Supplemental Table 2.

Detailed information is included in the Supplemental Material.

### Conflicts of interest

The authors have declared that no conflicts of interest exist.

### Acknowledgements

This work was supported by the National Research Foundation [2015R1C1A2A01051560 to M.J.K.] and the Asan Institute for Life Sciences [2018-577 to M.J.K.].

## Figure Legends

**Fig. 1. The changes in expression of metabolic genes in *fliI* mutants.** Quantitative RT-PCR analysis of each gene in seven-day-old *w<sup>1118</sup>* flies (controls) and *fliI<sup>3/14</sup>* mutants. The values were normalized to *Rp49* data. (A) Relative mRNA abundance of glycolytic-enzyme genes. (B) Relative mRNA expression of TCA cycle genes. (C) Relative expression of *Pgd* mRNA. (D) Relative expression levels of lipogenic genes. (E) Relative mRNA expression of lipolytic genes. Data are presented as mean  $\pm$  SE from at least five independent experiments; \**p* < 0.05, \*\**p* < 0.01, and \*\*\**p* < 0.001.

**Fig. 2. *FliI* mutants show increased levels of precursors of long-chain fatty acids.** The metabolites from seven-day-old *w<sup>1118</sup>* flies (controls) and *fliI<sup>3/14</sup>* mutants were monitored by LC-MS/MS. (A) The amounts of intermediates of glycolysis. The values of each metabolite amount were normalized to the total protein level. (B) The relative levels of intermediates of the TCA cycle. These amounts significantly increased in *fliI<sup>3/14</sup>* mutants compared to *w<sup>1118</sup>* flies, the controls. (C) The levels of intermediates of the pentose phosphate pathway (PPP). The ratio of NADPH/NADP involving a byproduct of the PPP, which was strongly diminished in *fliI<sup>3/14</sup>* mutant flies. The relative levels of fatty acyl CoAs are illustrated in (D). The amounts of precursors of long-chain fatty acyl-CoAs, such as palmitoyl-CoA, stearoyl-CoA, and oleoyl-CoA, were considerably larger in *fliI<sup>3/14</sup>* mutants relative to *w<sup>1118</sup>* flies (controls). Data are presented as mean  $\pm$  SE from at least three independent experiments; \**p* < 0.05, \*\**p* < 0.01, and \*\*\**p* < 0.001.

**Fig. 3. A schematic diagram of glucose metabolism and lipid metabolism according to the data from Figs. 1 and 2.** The red color indicates that mRNA or metabolite levels were

significantly upregulated in terms of statistics in *fliI*<sup>3/14</sup> mutants, the blue color denotes downregulation in *fliI*<sup>3/14</sup> mutants, and black indicates no change.

**Table 1. GO terms of genes regulated in *flightless-1* mutant flies.**

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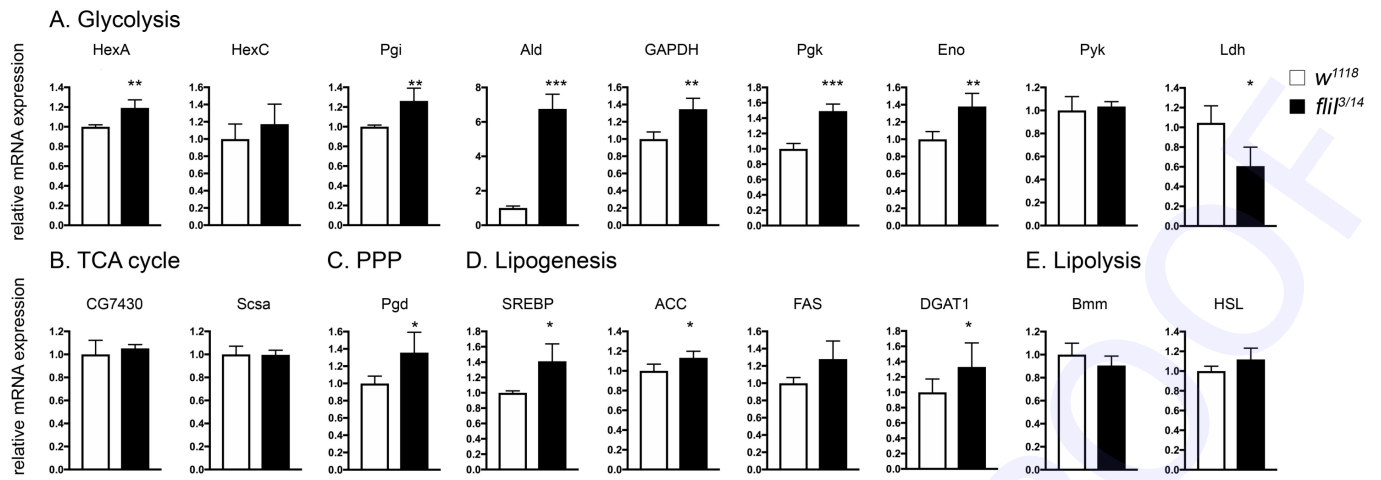
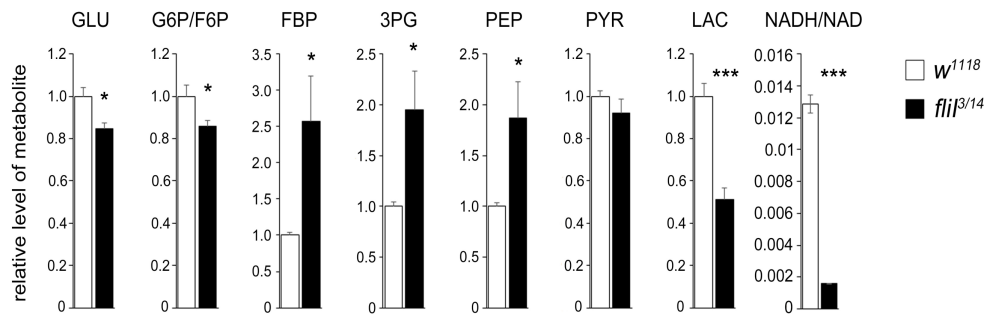


Fig. 1

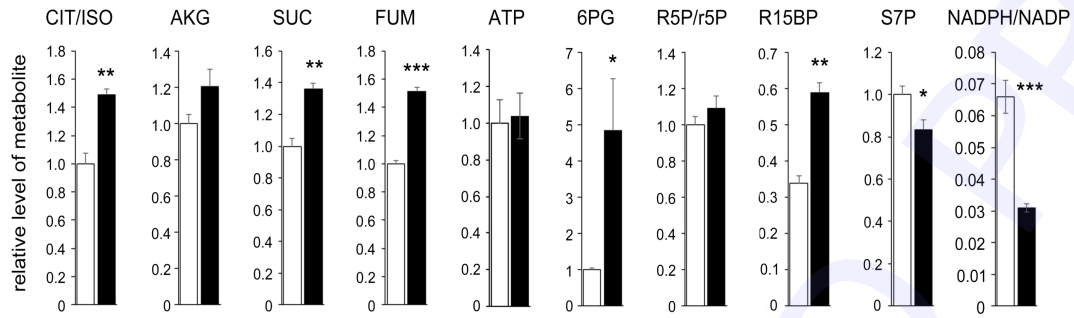
Fig. 1. Figure 1



## A. Glycolysis



## B. TCA Cycle



## C. Pentose Phosphate Pathway

## D. Fatty acyl CoA

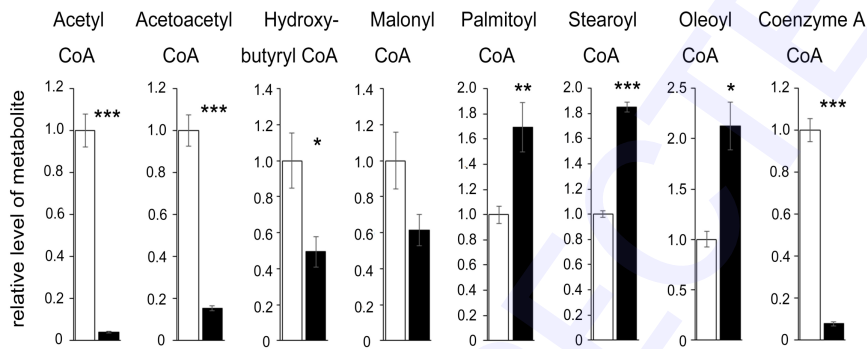


Fig. 2. Figure 2

Fig. 3. Figure 3

Table 1. GO terms of genes regulated in *flightless-1* mutant flies

*downregulated*

GO term	P value	Benjamini	Genes
GO:0051276 - chromosome organization	$3.98 \times 10^{-7}$	$7.44 \times 10^{-5}$	CG33822, CG33810, CG33825, RHI, CG33816, CG33828, CG33861, CG33831, CG33858
GO:0031057 - negative regulation of histone modification	$8.97 \times 10^{-7}$	$8.39 \times 10^{-5}$	CG33822, CG33810, CG33825, CG33816, CG33828, CG33831
GO:0006334 - nucleosome assembly	$1.58 \times 10^{-6}$	$9.87 \times 10^{-5}$	CG33822, CG33810, CG33850, CG33825, CG33816, CG33828, CG33861, CG33844, CG33831, CG33858, CG33847
GO:0005975 - carbohydrate metabolic process	$2.70 \times 10^{-6}$	$1.26 \times 10^{-4}$	MAL-A7, MAL-A6, TOBI, MAL-A8, AMY-P, MAL-A1, AMY-D, AMYREL, CHT8, MAL-B1
GO:0006342 - chromatin silencing	$1.39 \times 10^{-4}$	$5.19 \times 10^{-3}$	CG33822, CG33810, CG33850, CG33825, CG33816, CG33828, CG33844, CG33831, CG33847

*upregulated*

GO term	P value	Benjamini	Genes
GO:0006508 - proteolysis	$3.74 \times 10^{-9}$	$7.29 \times 10^{-7}$	CG9897, CG32523, ANCE-4, SPH93, CG42694, CG8539, CG1304, CG17239, SER6, CG15254, CG11842, GAMMATRY, CG9676, CG31267, CG7829, CG31681, CG3088, CG14529, ZETATRY, CG17475, CG8329, CG11911
GO:0050830 - defense response to Gram-positive bacterium	$1.53 \times 10^{-5}$	$1.49 \times 10^{-3}$	LYSP, LYSE, LYSD, SPH93, LYSB, GNBP1, IM23
GO:0019835 - cytolysis	$9.95 \times 10^{-5}$	$6.45 \times 10^{-3}$	LYSP, LYSE, LYSD, LYSB
GO:0016998 - cell wall macromolecule catabolic process	$2.32 \times 10^{-4}$	$1.13 \times 10^{-2}$	LYSP, LYSE, LYSD, LYSB

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## Supplemental material and methods

### Metabolic profiling

Fruit flies were homogenized using TissueLyzer (Qiagen) with MeOH. Internal standard solutions (malonyl- $^{13}\text{C}_3$  CoA, 5  $\mu\text{M}$  Gln- $\text{d}_4$ ) were added to the samples. Then, the samples were centrifuged at 13 000 rpm for 10 min (Eppendorf Centrifuge 5415R). The precipitate was stored for measurement of protein amounts later by the Bradford assay. For the supernatant, the aqueous phase after liquid-liquid extraction was collected, and used for analysis. Metabolites were analyzed on LC-MS/MS (1290 HPLC (Agilent)-Qtrap 5500 (ABSciex)). For metabolites related to energy metabolism, Synergi Fusion RP  $50 \times 2$  mm was employed. Here, 5 mM  $\text{CH}_3\text{COONH}_4$  in  $\text{H}_2\text{O}$  and in MeOH served as mobile phases A and B, respectively. The separation gradient was as follows: hold at 0% B for 5 min, 0% to 90% B for 2 min, hold at 90% for 8 min, 90% to 0% B for 1 min, and then hold at 0% B for 9 min. LC flow was 70  $\mu\text{l}/\text{min}$ , except for 140  $\mu\text{l}/\text{min}$  between 7-15 min, at 23  $^\circ\text{C}$ . For fatty acyl CoAs, a Zorbax 300 Extend-C18 column ( $2.1 \times 150$  mm) was used. Mobile phase A was  $\text{ACN}-\text{H}_2\text{O}$  (10:90) with 15 mM  $\text{NH}_4\text{OH}$ , and mobile phase B was ACN containing 15 mM  $\text{NH}_4\text{OH}$ . The separation gradient was the following: hold at 0% B for 3 min, 0% to 50% B for 2 min, 50% to 80% B for 5 min, 80% to 0% B for 0.1 min, and then hold at 0% B for 4.9 min. LC flow was 200  $\mu\text{l}/\text{min}$ , and column was kept at 25  $^\circ\text{C}$ . Multiple reaction monitoring (MRM) was employed for analysis. The quantitative value of each metabolite was normalized to the total protein amount.

### Transcriptome sequencing

Total RNA from the flies was isolated using the Trizol reagent (Invitrogen, USA). A library was prepared with 1  $\mu\text{g}$  of total RNA for each sample by Illumina TruSeq mRNA Sample

Prep kit (Illumina, Inc., San Diego, CA, USA). The libraries were quantified using qPCR according to the qPCR Quantification Protocol Guide (KAPA Library Quantification kits for Illumina Sequencing platforms) and qualified using the 2100 Bioanalyzer (Agilent Technologies, Waldbronn, Germany). Indexed libraries were then sequenced using the HiSeq4000 platform (Illumina, San Diego, USA by the Macrogen Incorporated).

### **Read mapping and differential gene expression analysis**

Reads were filtered with NGS QC Toolkit (1) (v.2.3.3) and mapped to the *D. melanogaster* reference transcriptome assembly (dm6) with Bowtie (2) and RSEM (3). Differential gene expression analysis was carried out with expected counts data from RSEM results using DEseq2 (4). We adjusted all p-values using multiple testing with a Benjamini-Hochberg correction with a false discovery rate (FDR) of 5%. Differentially expressed genes were identified with following thresholds: a fold change > 2, a p-value < 0.05 and an adjusted p-value < 0.05.

### **Enrichment analysis**

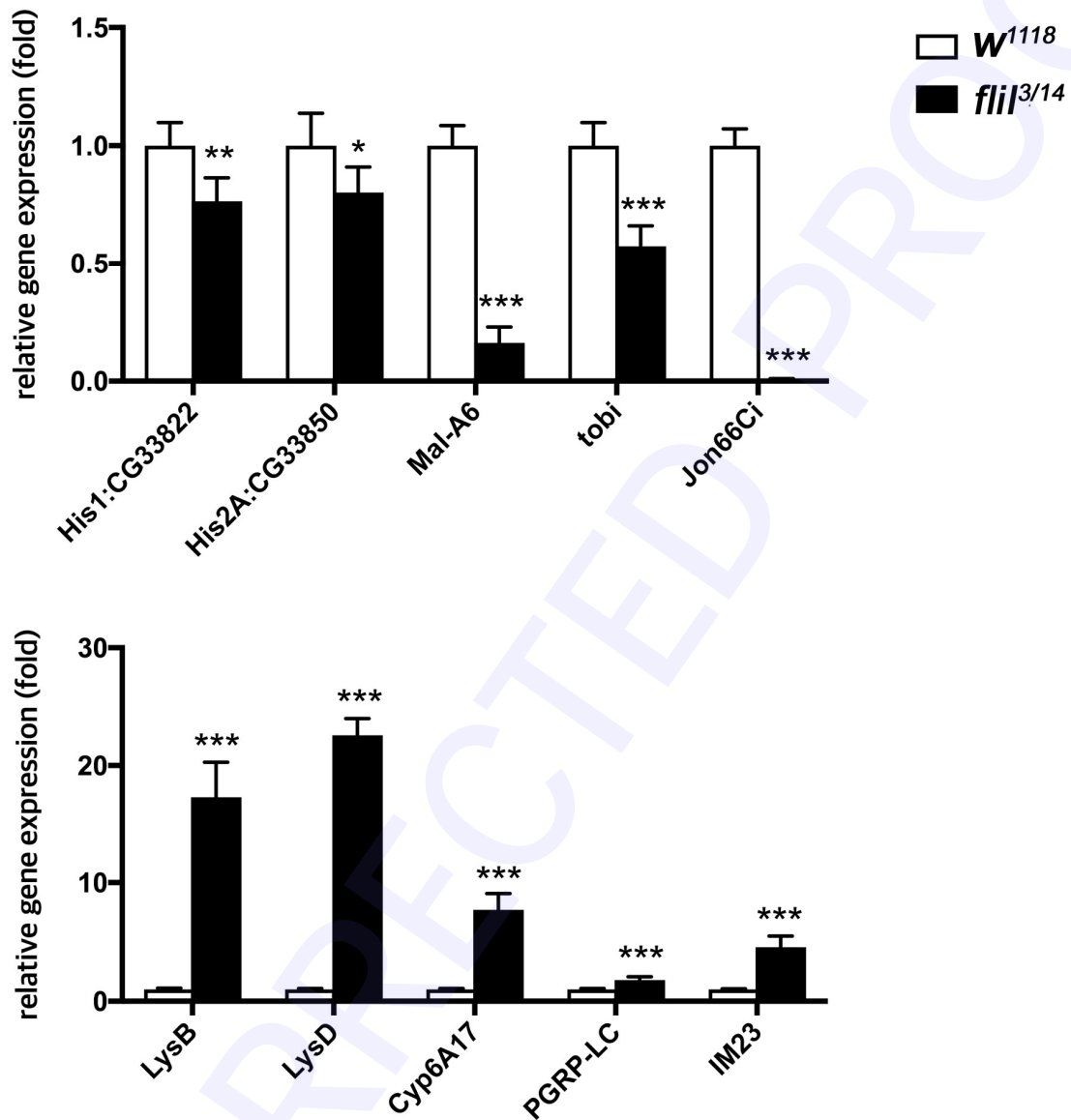
Identification of GO biological processes related to genes characterized with significant expression change was conducted using Database for Annotation, Visualization and Integrated Discovery (DAVID) (5). Significant GO biological processes were defined by p-value < 0.05 and FDR < 0.05.

### **Statistical analysis**

Each experiment was repeated at least three times, and the data are presented as mean  $\pm$  SE. Significance testing was based on Student's *t* test.

## References

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4. Love, M. I., Huber, W. and Anders, S. (2014) Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol* **15**, 550.
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**Supplemental Figure 1. Quantitative real-time PCR analysis for representative gene expression changes in *w<sup>1118</sup>* and *fliI<sup>3/14</sup>* mutants.** The values were normalized to rp49 levels.

Data are presented as mean  $\pm$  SE from at least three independent experiments; \* $p < 0.05$ , \*\* $p < 0.01$ , and \*\*\* $p < 0.001$ .



Supplementary Table 1

DEG_list_high						
gene_id	baseMean	log2FoldChange	lfcSE	stat	pvalue	padj
CG43090 12798122	385.7350469	12.17355009	1.495046108	8.142591737	3.87E-16	7.23E-14
p24-2 318890	354.9582182	12.05286279	1.475049448	8.171161796	3.05E-16	5.91E-14
CG1304 33074	68.31774398	9.674951082	1.570668563	6.159766171	7.29E-10	6.64E-08
CR40469 5740321	36.78760781	8.783916674	1.644660137	5.340870418	9.25E-08	5.97E-06
Lysd1 38127	1633.068465	8.635257928	0.878140805	9.833568694	8.07E-23	2.87E-20
CG32581 318098	571.7986249	7.96403544	1.955580493	4.07246619	0.00151326	0.05504785
CR46123 26067439	15.02179023	7.491178964	1.807571948	4.144332387	0.001174287	0.001174287
CG13032 399663	30.47718627	7.049831905	1.592032778	4.428195198	9.50E-06	0.000380121
CG45704 1306	1406.394415	6.779828363	0.248792211	27.25096711	1.62E-163	1.78E-159
CG44142 14462464	8.23406377	6.623349329	2.068456756	3.202072902	0.001364425	0.025455408
CG15534 34613	782.7497285	6.60024692	1.532013037	4.308217055	1.65E-05	0.000617215
CR43426 12798470	7.683276003	6.523486519	2.108159429	3.094399043	0.00197212	0.033922197
nrs1 37577	17.06427986	6.283834203	2.041647844	3.07782247	0.002085375	0.035465384
Cyp4p2 35946	47.70624482	6.095091485	1.20129024	5.073787567	3.90E-07	2.22E-05
CG14456 40481	15.44482987	6.088590278	1.850818521	3.289674385	0.001003034	0.02029257
Lys8 38125	374.1279326	5.60840238	1.505983972	3.7240784	0.00019603	0.005403567
Cyp6d2 37594	140.3247031	5.443756137	0.89003933	6.116309642	9.58E-10	8.45E-08
Cyp6a17 45556	209.7293961	5.318287221	0.509717624	10.43379112	1.74E-25	7.67E-23
Cyp309a2 33439	116.0927709	5.282468044	0.937878294	5.632359846	1.78E-08	1.29E-06
SPH9 35049	47.43717378	5.278225958	1.716963417	3.074163319	0.002110938	0.035569279
Oyr9b 43497	321.819425	5.264764956	1.263525075	4.16672772	3.09E-05	0.001074785
CG33337 2768678	26.56681867	5.23699243	1.100727508	4.757755569	1.96E-06	9.22E-05
CG32212 317917	120.4036977	5.201175623	0.524145724	9.441748093	3.30E-23	1.21E-20
SG2 3424	62.3675854	5.005508249	0.843609337	5.833443397	2.97E-09	1.76E-0708
CG30382 246582	793.0894746	4.906466706	0.377010906	13.01460152	1.01E-38	9.29E-36
CG14715 41403	549.574085	4.883335197	0.294277003	16.594387	7.66E-62	2.11E-58
GstE1 37106	591.7204962	4.676472047	0.290906549	16.07551313	3.79E-58	5.97E-55
CG4195 37018	309.957321	4.475154517	0.286172009	15.67398825	4.01E-55	4.92E-52
CG15711 36834	14.86883953	4.457889284	1.455651741	3.062469654	0.00219587	0.036506992
CG35509 3346225	15.16901876	4.331014639	1.449232609	2.988488261	0.002803613	0.043538921
Hesr1 32800	80.51000781	4.233321809	0.527082761	8.031607407	9.62E-16	1.74E-13
gammaTry 36221	687.5566106	4.095240571	1.204177667	3.400860758	0.00067174	0.014460333
Imp21 8674044	38.50697147	3.997650475	0.757891133	5.274702792	1.33E-07	8.33E-06
DM23 37099	38.17373541	3.959791435	1.300123261	3.045704629	0.00231358	0.038031632
CG32641 318136	111.7956183	3.928411629	0.473857584	8.290279103	1.13E-16	2.35E-14
Lys6 38128	268.0427113	3.778601592	0.613920655	6.154868626	7.51E-10	6.79E-08
SG12 49814	18.98749814	3.753735389	1.08968335	3.44474649	0.00189408	0.02189408
CG12653 33004	66.14360973	3.701318809	1.01731332	3.700209995	0.000215421	0.005835893
CG17470 35321	97.5859598	3.747874038	0.477464185	7.84953962	4.18E-15	7.19E-13
Unc-115c 41178	600.6922525	3.720074578	0.859869549	4.326324363	1.52E-05	0.000572514
vis 36372	105.8914909	3.680582968	0.559422607	6.579253188	4.73E-11	6.493055551
CG32594 41739	72.27370258	3.62379493	0.612474106	5.916469563	3.29E-09	2.68E-07
CG8768 36400	204.9775998	3.605685493	0.307704214	11.7180244	6.31E-29	2.96454269
CG18934 3056	48.97910361	3.561276636	0.70066686	5.082695985	3.72E-07	2.13E-05
sosie 42961	600.9565065	3.423107401	0.208692914	16.60260315	1.83E-60	4.04E-57
CG10182 42780	63.71532738	3.405772905	0.761725121	4.471131336	7.78E-06	0.000318922
Cyp6a14 35835	57.29817113	3.398101639	0.758986329	4.471579902	7.56E-06	0.000312376
CG15254 34915	156.4514089	3.313522797	0.420683283	7.86259947	3.37E-15	5.99E-13
CG34423 5740313	135.7439029	3.219752827	0.453937777	6.079238696	1.37E-10	1.77E-10
CG2064 35708	234.4982022	3.138966094	0.489405651	6.413832962	1.42E-10	1.37E-08
Unc-43 5498	129.1005562	3.070136436	0.30308646	3.39643087	0.000674835	0.014504346
CG59614 151541	467.8178785	2.976941687	0.21823272	13.64115548	2.28E-42	2.28E-39
Act88f 41885	256.7003012	2.972416658	0.438816865	6.737706509	1.26E-11	1.40E-09
Ipod1 31954	27.6267349	2.970495903	0.77011448	3.857213415	0.000114687	0.003417673
Hsp70 8b 50022	136.3041967	2.954080208	0.97769703	3.021467639	0.002515523	0.003401481
Cyp6a2 35587	391.8760965	2.910802197	0.450095932	6.467070662	9.99E-11	9.93E-09
Tap4 35248	1084.251408	2.895487398	0.302518898	9.571261228	1.06E-21	3.53E-19
CG55915 43765	3556.397689	2.886395425	0.156968838	18.38833398	1.63E-75	5.99E-72
Unc 34060	474.2833766	2.878713551	0.406359985	7.084146219	1.40E-12	1.86E-10
phr6-4 35322	316.0475746	2.828328133	0.252999034	11.17920527	5.15E-29	2.96E-26
CG3819 40069	225.2015691	2.725827882	0.474284392	5.747243487	9.07E-09	6.95E-07
CG34141 4379870	173.1971671	2.723474382	0.312521414	8.714520871	2.92E-18	6.85E-16
Uhg5 8673975	145.0524012	2.720028781	0.412142787	6.599724332	4.12E-11	4.28E-09
CG55688 36658	555.5949254	2.64524801	0.346356978	7.63734759	2.22E-14	3.70E-12
CG33237 2768939	55.49837988	2.549837988	0.30308646	3.39643087	0.000674835	0.014504346
CG14692 41298	61.19092474	2.549219275	0.547747814	4.65400712	3.26E-06	0.000145326
Ilp7 71328	36.86704764	2.543225088	0.814324223	3.123111184	0.001789501	0.031243189
CR43973 14462864	37.59819688	2.369092174	0.630379349	3.758200769	0.00017114	0.004863362
Cyp4p3 35948	67.59411115	2.246636409	0.674821999	3.32928189	0.00087087	0.017914582
CG18343 36297	130.2222071	2.20170765	0.2310391	9.529588934	1.58E-21	4.84E-19
CG15126 50202	26.35439874	2.193754046	0.724129947	3.029503277	0.002449562	0.03958573
CG45071 19834908	155.0380108	2.191164379	0.287535437	7.62052031	2.53E-14	4.16E-12
w 31271	93.15242074	2.180097842	0.417848586	5.217435016	1.81E-07	1.12E-05
pncr014.3 10178937	147.0957588	2.172110689	0.34977889	6.209953642	5.30E-10	4.87E-08
CG31681 318882	460.4417462	2.163306573	0.448613324	4.822207578	1.42E-06	6.93E-05
CG8539 38842	74.94327742	2.160377242	0.454194641	4.756500951	1.97E-06	9.24E-05
CG9676 32647	759.5410725	2.1603192	0.353798118	6.433386861	1.25E-10	1.22E-08
CG32243 317935	699.193622	2.117025193	0.171078956	12.37455057	3.59E-35	3.32E-32
Ilp2 3194	300.0508728	2.106237866	0.25440202	8.18146255	0.000145326	0.014504346
CG14691 41982	215.2395326	2.088565175	0.306177423	6.821421011	1.01E-12	1.01E-09
CG7408 39991	176.77322	2.074113348	0.360269735	5.757112617	8.56E-09	6.64E-07
CG13091 34188	564.1296426	2.04423976	0.302261395	6.763151961	1.35E-11	1.47E-09
CG78294 43522	91.89434032	2.043630502	0.553486317	3.692287307	0.00022246	0.00597696
Cyp12d1-p 264648	176.9877362	1.995869142	0.416913881	4.7872456	0.00757196	0.0807573
CG1397 46386	3442.816471	1.94617562	0.105670796	18.41734616	9.54E-76	5.26E-72
Ser6 33073	140.260446	1.886357901	0.51579974	3.657151711	0.000255033	0.006743402
PGRP-LC 39063	680.1529209	1.878143693	0.227623119	8.251111323	1.57E-16	3.15E-14
CG11211 35545	92.98517366	1.844055911	0.473714748	3.892755961	0.003027167	0.030027167
CG30404 35496	1387.796765	1.836199908	0.165673024	11.08327637	1.51E-28	8.34E-26
CG8329 39123	827.9202743	1.79789854	0.259753633	6.921904551	4.46E-12	5.52E-10
CG11842 43431	70.2704535	1.783848364	0.392703459	4.216383459	0.00085832	0.013860582
CG33748 377288	50.15412306	1.73734905	0.53136708	3.111237306	0.00112376	0.011237306
CG32453 3797953	66.26323775	1.73249878	0.490578872	6.23444848	0.000144848	0.00144848
CG15529 43584	205.7281732	1.728795793	0.389243507	4.44142887	8.94E-06	0.0003693
CG14529 43408	86.19729379	1.718759614	0.576906252	2.979270217	0.002889359	0.044247315
CG32708 318161	237.6263576	1.705806575	0.266547105	6.8989578	1.56E-10	1.49E-08
CG3104 33486	58.6888752	1.70565964	0.5462608	3.124247309	0.001793664	0.031243189
CG17239 59224	254.3356147	1.693401406	0.425490115	3.979884247	6.89E-05	0.002190964
CG12105 38190	78.7541879	1.680580676	0.454732602	3.69575585	0.000219234	0.005910194
CR45873 26067201	59.92574281	1.67849902	0.556110874	3.004526577	0.002659945	0.042017988
Uhg134 344	426.3881344	1.662404701	0.330484381	5.030206561	4.90E-07	2.71E-05
CG4302 37420	382.9207183	1.643917114	0.32833994	5.006753417	5.54E-07	2.95E-05
simj 39212	1889.376435	1.639149933	0.176873136	9.26737667	1.91E-20	5.01E-18
Ance-4 35909	385.5436808	1.615410197	0.535576658	3.016027245	0.002595983	0.040855436
CG5621 42476	232.85462	1.60702193	0.355099625	4.525552316	6.02E-06	0.000257643
Smyd4-4 42034	10.7622216	1.571752603	0.452908695	3.470446068	0.000519605	0.012138066
Avx1 3819	220.0936124	1.15682583	0.156878276	2.96E-23	1.13E-20	1.13E-20
hnp3 38090	220.0936124	1.546468832	0.253659815	6.089600082	1.13E-09	9.91E-08
CR31781 12798248	1508.970537	1.543195836	0.178852937	8.62829465	1.23E-18	1.43E-15
CG3088 39115	768.3987474	1				



AOX3 41896	2216.845689	-1.090930283	0.107074883	-10.18847979	2.23E-24	9.12E-22
CG10170 42774	137.8484501	-1.09034477	0.351675857	-3.100425431	0.001932429	0.033448914
CG14102 40156	426.8384604	-1.083164177	0.196222855	-5.52007143	3.39E-08	2.35E-06
Chl8 37390	436.2128371	-1.041567481	0.210627684	-4.945064497	7.61E-07	3.89E-05
Glaz 36447	408.7136762	-1.04039084	0.197280021	-5.273675627	1.34E-07	8.33E-06
CR43837 14462385	533.5606538	-1.037115447	0.251407467	-4.125237254	3.70E-05	0.001260342
CG7565 38893	858.9272342	-1.036249054	0.176620293	-5.867100751	4.43E-09	3.57E-07
CG11961 37221	1214.955439	-1.035427226	0.275762275	-3.754782003	0.000173492	0.004904942
CG14036 33735	778.0974105	-1.02076211	0.178385279	-5.7222329	1.05E-08	7.99E-07
Cyp9f2 41520	1277.844993	-1.01873118	0.185202228	-5.50064214	3.78E-08	2.59E-06
Oatp58Dc 37545	542.3278022	-1.018041126	0.244963006	-4.155897425	3.24E-05	0.001119928
CR45835 26067163	201.9671134	-1.017252548	0.301658081	-3.372203869	0.000745692	0.015750964
CG4842 39817	464.9455976	-1.016533148	0.180979456	-5.616842753	1.94E-08	1.40E-06
Mco1 34258	186.5853803	-1.01589402	0.310187224	-3.275099497	0.001056248	0.021136455
Ir76a 40157	178.3480049	-1.015236821	0.319103254	-3.181530769	0.00146499	0.026921626
CG5096 34410	705.5381285	-1.010657036	0.267373074	-3.779950692	0.000156859	0.004527571
Gih 34927	420.5673888	-1.003827696	0.326183979	-3.077489275	0.002087524	0.035465384



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**Suppelmental Table 1. PCR primers used in this studies**

gene name	forward sequences	Reverse sequences
HexA	ATATCGGGCATGTATATGGG	CAATTTGCTCACATACTTGG
HexC	GGCTATACTCAACGATACCAC	CGCAATAGGTCCACATTCTC
Pgi	ACTGTCAATCTGTCTGTCCA	GATAACAGGAGCATTCTTCTCG
Ald	GGCAAGAAGGAGAACATTGC	CAACCAAACGCCTTAGTAGG
Gapdh	CTCGCATATAATCACGCGTC	CACCTTGCCATACTTCTTGTC
Pgk	CTGATTGAGAACCTTCTGGAC	CTTCTCCACCAGTTTCTCGA
Eno	CAACATCCAGTCCAACAAGG	GTTCTTGAAGTCCAGATCGT
Pyk	GTGCCACTCATCTACAAGGA	ATGAAGCCGTTCTTCTTTCC
Ldh	ACAAAACAGAGCGGAATGCT	CTACGATCCGTGGCATCTTT
CG7430	AGAGCGAGGAGCAACTGAAG	GTTGGTCTTGGCACGAGAGT
Scsα	CGTCATCTATGTGCCACCAC	ACACCCTCCGTAATGCAAAC
Pgd	TCGCCAATGAGGCTAAGGAC	CCAGCAGCATGACCTTCCG
SREBP	CGCAGTTTGTGCGCTGATG	CAGACTCCTGTCCAAGAGCTGTT
ACC	GTGCAACTGTTGGCAGATCAGTA	TTTCTGATGACGACGCTGGAT
FAS	GACTTGACCGATCCGATCAAC	CCCCAGGAGGTGAACTCTATCA
DGAT1	AGTCTGCGAAACCGCAAGTC	ATCCGTTGGTTCCATTACCATT
Bmm	GTCCCTTCAGTCCCTCCTTC	TATGAAGCACGCACACAACA
HSL	GTTGCGATGCGGAAATCACACTGC	GAGAACTCCGCGTATCGAGTCG
CG33822	CCCAAAAAGACGGTGAAGAA	TTTTTGGCAGCCGTAGTCTT
CG33850	TTCTGTTGCCCAAGAAGACC	AAAGGACGGTTTGATTGACG
Mal-A6	GACCTTGAGAGAGCGACCAC	TTGTAGGAGCGCGGATAGAT
tobi	GATGCAGCCATTTTGGACTT	ATGCCATCCTCATCCTGAAG
Jon66Ci	GCTTGTGGGAGTCACCAACT	GTCACGGATCCAGTCCAAGT
LysB	ACTACAACGGCTCCAACGAC	CTCAACCCGCACTCATTGTA
LysD	CTGGTCCACCTGGCACTACT	GGTGCGCAATCGTTTTAATC
Cyp6A17	TGCAACAGCTTGTACGATCC	AGCATATTCCCGTAGCGATG
PGRP-LC	TTTGTCTTTTCTGCCCAAC	TACGATACCCAGCAGGAAGG
IM23	GCACGCAGATTGAGAATGAA	CTCCGCCGATAATCACATT
rp49	AGATCGTGAAGAAGGCACCAAG	CACCAGGAACTTCTTGAATCCGG