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Promotion of formyl peptide receptor 1-mediated neutrophil chemotactic migration by antimicrobial peptides isolated from the centipede *Scolopendra subspinipes mutilans*

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Running title: New agonists for FPR1

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Keywords: antimicrobial peptide, neutrophil, formyl peptide receptor 1, chemotaxis, *Scolopendra subspinipes mutilans*

Abstract

We investigated the effects of two antimicrobial peptides (AMPs) isolated from *Scolopendra subspinipes mutilans* on neutrophil activity. Stimulation of mouse neutrophils with the two AMPs elicited chemotactic migration of the cells in a pertussis toxin-sensitive manner. The two AMPs also stimulated activation of ERK and Akt, which contribute to chemotactic migration of neutrophils. We found that AMP-stimulated neutrophil chemotaxis was blocked by a formyl peptide receptor 1 (FPR1) antagonist (cyclosporin H); moreover the two AMPs stimulated the chemotactic migration of FPR1-expressing RBL-2H3 cells but not of vector-expressing RBL-2H3 cells. We also found that the two AMPs stimulate neutrophil migration *in vivo*, and that this effect is blocked in FPR1-deficient mice. Taken together, our results suggest that the two AMPs stimulate neutrophils, leading to chemotactic migration through FPR1, and the two AMPs will be useful for the study of FPR1 signaling and neutrophil activation.

Introduction

Neutrophil migration is an important event in the immune response against infection or tissue damage (1). Tissue-resident sentinel cells such as macrophages and dendritic cells recognize and capture invading pathogens, resulting in the activation of the sentinel cells and the production of chemokines (2). The chemokines cause recruitment of circulating neutrophils in the blood to the infected site (3,4). In addition to chemokines, invading bacteria-derived molecules also cause the recruitment of neutrophils into infected sites (3,4). One important bacteria-derived neutrophil recruiting molecule is formyl peptide (5). As chemotactic neutrophil migration into sites of infection or injury is crucial in regulating immune or inflammatory responses, it is important to identify new molecules that can control neutrophil migration.

Scolopendra subspinipes mutilans has been used in oriental medicine to treat rheumatoid arthritis, lymphadenopathy, and carcinoma (6,7). However, the identities of the bioactive components that regulate the activity of leukocytes or other cell types have not been extensively studied. In a previous report, we identified some antimicrobial peptide (AMP) candidates through *de novo* RNA sequencing from *Scolopendra subspinipes mutilans* (8). We demonstrated that some of the AMPs have dual activity, with bactericidal activity and anti-cancer activity against leukemia (9). Recently we also reported that one AMP isolated from *Scolopendra subspinipes mutilans* stimulates macrophage chemotaxis (10). In this study, we asked whether AMPs isolated from *Scolopendra subspinipes mutilans* stimulate neutrophil activity. We found that two AMPs from *Scolopendra subspinipes mutilans* stimulate neutrophil activity. We found that two AMPs from *Scolopendra subspinipes mutilans* [scolopendrasin III (sequence:

VIYVHCTSSRGNNVRLLCASKVAP-NH₂), scolopendrasin V (sequence: YYGGGYKYKHWGCR-NH₂)], strongly stimulate neutrophil chemotaxis. We also revealed that neutrophil chemotaxis induced by the two AMPs is mediated by an important neutrophil chemoattractant receptor, formyl peptide receptor (FPR) 1.

Results

Two novel AMPs stimulate neutrophil chemotactic migration in a pertussis toxin (PTX)-sensitive manner

Previously, we and others demonstrated that AMPs, including LL-37 and scolopendrasin VII, stimulate chemotactic migration of leukocytes (10,11). In this study, we tested the effects of two new AMPs (scolopendrasin III and scolopendrasin V) isolated from *Scolopendra subspinipes mutilans* on neutrophil activity. Because the three scolopendrasin AMPs have antimicrobial activity, we compared the amino acid sequence between the three scolopendrasin AMPs using multiple sequence alignment with hierarchical clustering as described previously (12). We found that some amino acids are commonly conserved between two of the AMPs among three scolopendrasin AMPs (Fig. 1A). However, well-conserved amino acids were not detected among the three AMPs (Fig. 1A). We examined the effects of the two new AMPs on neutrophil chemotaxis using a Boyden chamber assay kit. Stimulation of mouse neutrophils with the two AMPs strongly elicited neutrophil chemotactic migration (Fig. 1B). AMPs-induced neutrophil chemotactic migration was concentration-dependent, and maximal activity was induced at 100 µg/ml AMP (Fig. 1B).

Many chemokines and chemoattractants induce neutrophil chemotaxis via PTX-sensitive

G-protein(s) (13,14). Therefore, we also investigated whether AMP-induced neutrophil chemotaxis is mediated by PTX-sensitive G-protein(s). Preincubation of neutrophils with PTX prior to the chemotaxis assay with the two AMPs almost completely abolished peptide-induced neutrophil chemotaxis (Fig. 1C). In a positive control experiment, we found that WKYMVm-induced neutrophil chemotaxis was also completely blocked by PTX treatment (Fig. 1C). The results indicate that the two AMPs stimulate neutrophil chemotactic migration via PTX-sensitive G-protein(s).

In addition to chemotactic migration into the infectious or injured area, neutrophils mediate immune responses by stimulating production of superoxide anion (15). We examined whether the two AMPs stimulate superoxide anion production from mouse neutrophils using cytochrome c reduction assay. As shown in Fig. 1D, both AMPs failed to stimulate superoxide anion production. However, the well-known FPR agonist WKYMVm strongly stimulated the production of superoxide anion from neutrophils (Fig. 1D). Degranulation is also important to mediate immune responses against infection (16). The two new AMPs did not stimulate degranulation in neutrophils (Fig. 1E).

Two new AMPs selectively stimulate intracellular signaling downstream of PTX-sensitive G-protein(s)

There are wide range of extracellular stimuli that elicit diverse intracellular signaling cascades, which mediate various cellular responses. Intracellular calcium increase is an important event, which is associated with neutrophil activation in response to extracellular stimuli (17). In this study, we examined the effects of the two AMPs on calcium signaling in mouse neutrophils. Neither of the AMPs stimulated intracellular calcium increase in

neutrophils (Fig. 2A). However, stimulation of mouse neutrophils with WKYMVm strongly elicited intracellular calcium increase (Fig. 2A). ERK is known to play crucial roles in the regulation of neutrophil activity (18). We tested the effects of the two AMPs on the activity of the ERK by monitoring phosphorylation of the enzymes. Stimulation of mouse neutrophils with the two AMPs markedly induced phosphorylation of ERK (Fig. 2B). Phosphorylation of ERK was transiently induced by the two AMPs, showing time-dependent manner. Scolopendrasin III-induced ERK phosphorylation was transient showing maximal activity at 2-5 min and returning to basal levels at 10 min (Fig. 2B left). Scolopendrasin V-induced ERK phosphorylation was apparent at 5-30 min after stimulation (Fig. 2B right). Because the two AMPs stimulated chemotactic migration of neutrophils, we tested the roles of ERK on AMP-induced neutrophil chemotaxis using specific inhibitor of ERK (PD98059). Pre-incubation of neutrophils with an ERK inhibitor (PD98059) prior to the chemotaxis assay almost completely inhibited peptide-induced chemotaxis (Fig. 2C). These results indicate that neutrophil chemotaxis induced by the two AMPs is mediated by the activity of ERK.

Akt has also been reported to play a key role in the regulation of cell migration (19). In our study, we tested whether the two AMPs stimulate Akt activity. Stimulation of mouse neutrophils with the two AMPs elicited Akt phosphorylation at 2 to 10 min after stimulation (Fig. 2B). Pretreatment of neutrophils with Akt inhibitor (MK-2206) or phosphatidylinositol 3-kinase (PI3K) (upstream molecule of Akt) inhibitor (LY294002) prior to the chemotaxis assay completely blocked peptide-induced neutrophil chemotaxis, indicating that AMP-induced chemotaxis is mediated by PI3K/Akt pathway (Fig. 2C).

FPR1 mediates neutrophil chemotaxis induced by the two AMPs

Our finding that the two AMPs stimulate neutrophil chemotaxis, which is completely inhibited by PTX, led us to test the possible role of a well-known neutrophil chemoattractant receptor, FPR1, which is a PTX-sensitive G protein-coupled receptor. At first we examined the effect of an FPR1 antagonist, cyclosporin H (CsH) (20), on neutrophil chemotaxis stimulated by the two AMPs. Neutrophil chemotaxis was almost completely blocked by CsH. CsH also completely inhibited fMLF-induced neutrophil chemotaxis (Fig. 3A). To confirm that the AMP-induced neutrophil chemotaxis is mediated by FPR1, we used stably transfected FPR1-expressing RBL-2H3 cells (21). First, we used flow cytometric analysis to confirm that FPR1-transfected RBL-2H3 cells, but not the vector-transfected RBL-2H3 cells, express FPR1 on the cell surface (Fig. 3B). Next, we tested the effect of the two AMPs on chemotactic migration in vector- or FPR1-expressing RBL-2H3 cells. The two AMPs markedly stimulated chemotactic migration of cells in FPR1-expressing but not vector-expressing RBL-2H3 cells (Fig. 3C). The well-known FPR1 agonist fMLF also strongly induced the chemotactic migration of FPR1-, but not of the vector-expressing RBL-2H3 cells (Fig. 3C). However, neither of the AMPs stimulated the chemotactic migration of cells in the FPR2-expressing RBL-2H3 cells (Fig. 3D). The known FPR2 agonist MMK-1 significantly stimulated the chemotactic migration of FPR2-expressing RBL-2H3 cells (Fig. 3D). These results suggest that FPR1 but not FPR2 mediates the neutrophil chemotaxis induced by the two AMPs. Since the two new AMPs stimulate neutrophil chemotaxis via FPR1, we also investigated whether the two AMPs induce the chemotactic migration of macrophages, which also express FPR1. As expected, the two AMPs significantly stimulated the chemotactic migration of macrophages (Fig. 3E).

The two AMPs elicit neutrophil recruitment *in vivo* via FPR1

Since we observed that the two AMPs stimulate neutrophil chemotaxis *in vitro*, we then tested their effect on recruitment of neutrophils *in vivo*. Vehicle administration did not induce neutrophil recruitment into the peritoneal cavity, and no significant difference was observed in either WT or FPR1-deficient mice (Fig. 4A). Administration of the two AMPs into the peritoneal cavity strongly induced recruitment of neutrophils into the peritoneal cavity (Fig. 4B, 4C). Because we found that the two AMPs stimulate neutrophil chemotaxis through FPR1 at the cellular level, we also examined the functional role of FPR1 on AMP-induced *in vivo* neutrophil recruitment using FPR1-deficient mice. Neutrophil recruitment into the peritoneal cavity induced by the two AMPs was markedly attenuated in FPR1-deficient mice (Fig. 4B-D). These results indicate that *in vivo* recruitment of neutrophils toward the two AMPs is mediated by FPR1.

Discussion

In this study, we demonstrated that two novel AMPs stimulate neutrophil chemotactic migration, which is an important part of the innate immune system. We found that neutrophil migration stimulated by the two AMPs is mediated by FPR1 *in vitro* and *in vivo*. Since the AMPs stimulated neutrophil chemotaxis in FPR1-expressing RBL2H3 cells but not in FPR2-expressing RBL-2H3 cells (Fig. 3C and 3D), the two AMPs seem to be FPR1-selective agonists. Several previous reports demonstrated that various peptide agonists derived from hosts as well as bacteria bind to FPR1, resulting in the activation of

leukocytes (22,23). Such agonists include several formyl peptides and annexin 1 (23,24). However, evidence of AMP binding to FPR1 is limited. We previously demonstrated that one AMP derived from *Scolopendra subspinipes mutilans*, Scolopendrasin VII, binds to FPR1, resulting in macrophage chemotactic migration (10). Although the concentrations needed to activate FPR1 are extremely high for the two new AMPs compared with the already known FPR1 agonists such as WKYMVm (Fig. 1), our finding on the selective activation of FPR1 by the two other AMPs leading to neutrophil chemotaxis extends the concept that FPR1 is involved in the effect of AMPs on the activation of phagocytic leukocytes, such as macrophages and neutrophils.

Based on our previous report (10) and current finding on the action of two scolopendrasin AMPs (scolopendrasin III and scolopendrasin V), we compared their primary sequences. As shown in Fig. 1A, we could not detect any well-conserved amino acids among the three scolopendrasin AMPs. None of the three scolopendrasin AMPs has any amino acid sequences of the two well-known FPR1 agonists (fMLF or WKYMVm). The results suggest that other unrevealed factors other than the primary amino acid sequences might be involved in the binding of the AMPs to FPR1. Because the scolopendrasin AMPs were identified by analyzing the physiochemical properties including length (≤ 50 mer), pI ($8 \leq pI \leq 12$) and positive charge (8), further studies are needed whether or not these characteristics are essential for the binding of the AMPs to FPR1.

Since neutrophils play an important role in host immune defense by generating reactive oxygen species (e.g. the superoxide anion) and releasing granule contents, we tested the effects of the two novel AMPs on the generation of the superoxide anion and degranulation. Although the two AMPs strongly increased chemotactic migration of neutrophils, they did

not stimulate superoxide anion production; however, a well-known neutrophil stimulant, WKYMVm, strongly increased superoxide anion production from neutrophils (Fig. 1C). The two AMPs also failed to induce degranulation from neutrophils (Fig. 1D). Many FPR1 agonists have been reported to stimulate intracellular calcium increase, mediating diverse cellular responses (25,26). We investigated whether the two AMPs stimulate intracellular calcium increase using Fura-2/AM, a calcium binding dye. Stimulation of fura-2/AM-loaded neutrophils with the two AMPs did not increase intracellular calcium levels (Fig. 2A). However, a well-known neutrophil stimulant, WKYMVm, strongly induced intracellular calcium increase in the cells (Fig. 2A). Taken together, our results show that the two novel FPR1 agonists stimulate PTX-sensitive G protein/ERK/Akt pathways, leading to chemotactic migration of neutrophils, without affecting intracellular calcium release and subsequent degranulation and superoxide anion production from neutrophils. In a previous report, we demonstrated that FPR1 is differentially activated by different agonists in a ligand-selective manner (21). Unlike WKYMVm, the WKGMVm and WKRMVm stimulated phosphorylation of ERK/Akt and the subsequent chemotactic migration, but did not stimulate calcium increase and subsequent degranulation activity in FPR1-expressing RBL-2H3 cells (21). Considering that superoxide anion production and degranulation are mediated by intracellular calcium increase in neutrophils (27,28), our finding suggests that the activation of FPR1 by the two new AMPs do not induce calcium increase-mediated superoxide anion production and degranulation in the cells. The two new AMPs stimulate FPR1, showing selective activation of FPR1 downstream signaling leading to ERK/Akt phosphorylation and subsequent chemotactic migration. Previously, it has been suggested that one G-protein coupled receptor can be activated by different ligands via

ligand-selective receptor activation states (29). Binding of different ligands to one G-protein coupled receptor may induce distinct receptor conformation change, resulting in coupling of the receptor to different heterotrimeric G-proteins, and subsequent activation of different effector enzymes (29). Studies on the detail mechanism involved in the differential activation of FPR1 by the two new AMPs or WKYMVm are needed. In conclusion, in this study we report that the two AMPs (scolopendrasin III and scolopendrasin V) are novel FPR1 agonists. The two AMPs can be used as biased agonists for the selective activation of FPR1 downstream signaling.

Materials and Methods

Materials

Scolopendrasin III, scolopendrasin V, WKYMVm, and MMK-1 were synthesized by Anygen (Gwangju, Korea) with a purity > 99.6 %. fMLF and CsH were purchased from Enzo Life Sciences, Inc. (Farmingdale, NY, USA). Boyden chambers were purchased from Neuroprobe, Inc. (Gaithersburg, MD, USA). Antibodies against phosphorylated kinases including anti-phospho-Akt, anti-phospho-ERK were purchased from Cell Signaling Technology (Beverly, MA, USA). PD98059 was purchased from Calbiochem (San Diego, CA, USA). LY294002 was from BIOMOL Research Laboratories, Inc. (Plymouth Meeting, PA, USA). MK-2206 and *p*-nitrophenyl-*N*-acetyl- β -D-glucosamide were purchased from Sigma (St. Louis, MO, USA). Fura-2 pentaacetoxymethylester (fura-2/AM) was purchased from Molecular Probes (Eugene, OR, USA). RPMI 1640 was obtained from Welgene (Gyeongsan, Korea). FITC-conjugated anti-human FPR1 mouse IgG was purchased from

R&D Systems, Inc. (Minneapolis, MN, USA). Anti-Ly6G and anti-CD11b antibodies were obtained from e-Bioscience. Inc. (San Diego, CA, USA).

Isolation of mouse neutrophils

Mouse bone marrow neutrophils were isolated as previously described (30). Briefly, bone marrow cells isolated from femurs and tibias were suspended in HBSS-EDTA solution. After centrifugation at 400g for 10 min, resuspended cells were carefully loaded on a 52%/69%/78% Percoll gradient and centrifuged at 1500g for 30 min. Cells were isolated in the 69%/78% interface layer, then RBCs were removed by hypotonic lysis. Isolated cells were over 95% Ly6G-positive as assessed by flow cytometry (BD FACSCanto II).

Chemotaxis assay

Chemotaxis assays with mouse bone marrow neutrophils and vector-, FPR1-, or FPR2-expressing RBL-2H3 cells were performed in accordance with a previous report using a multiwell Boyden chamber (31). Cells were applied to different-sized polycarbonate filters (3 μ m pore size for mouse neutrophils, 5 μ m pore size for mouse bone marrow-derived macrophages, and 8 μ m pore size for the RBL-2H3 cells) for 90 min (2 h for the macrophages, 4 h for the RBL-2H3 cells) at 37°C. After removing non-migrated cells, the migrated cells were stained with hematoxylin (Sigma, St. Louis, MO, USA) and then quantified using a light microscope as previously described (31).

Measurement of superoxide anion production

Superoxide anion generation was determined by measuring reduction of cytochrome c

using a microtiter 96 well plate ELISA reader (Bio-Tek instruments, EL312e, Winooski, VT, USA) as described (31). Isolated mouse neutrophils (1×10^6 cells/100 μ l of RPMI 1640 medium per well of a 96-well plate) were stimulated with peptides in the presence of 50 μ M cytochrome c and 5 μ M cytochalasin B. The superoxide generation was measured as change in light absorption at 550 nm over 5 minutes at 1 min intervals.

Measurement of degranulation activity

Degranulation activity was determined by measuring the amount of released β -hexosaminidase as described previously (21). Isolated mouse neutrophils (1×10^6 /well) were washed twice with Tyrode's buffer (137 mM NaCl, 12 mM NaHCO₃, 5.6 mM glucose, 2.7 mM KCl, 1 mM CaCl₂, 0.5 mM MgCl₂, 0.4 mM NaH₂PO₄, 0.1g/100 ml BSA, and 25 mM HEPES, pH 7.4) and stimulated with individual peptides. The reaction was terminated 30 min after stimulation by placing the plate on ice. The amount of β -hexosaminidase secreted into the medium was determined by incubating supernatant or cell lysate using 5 mM *p*-nitrophenyl-*N*-acetyl- β -D-glucosamide in 0.1 M sodium citrate buffer (pH 3.8).

Measurement of intracellular calcium concentration

The level of intracellular calcium concentration was measured by Grynkiewicz's method using fura-2/AM (32). Isolated mouse neutrophils were incubated with 3 μ M fura-2/AM at 37°C for 50 min in fresh serum free RPMI 1640 medium under continuous stirring. 1×10^6 cells were aliquoted for each assay in Ca²⁺ free Locke's solution (154 mM NaCl, 5.6 mM KCl, 1.2 mM MgCl₂, 5 mM HEPES, pH 7.3, 10 mM glucose, and 0.2 mM EGTA). The

fluorescence changes at the dual excitation wavelengths of 340 nm and 380 nm and the emission wavelength of 500 nm were measured.

Measurement of surface expression of FPR1

Vector- or FPR1-expressing RBL-2H3 cells were incubated with FITC-conjugated monoclonal antibodies against FPR1 or isotype control for 40 min, followed by washing with FACS buffer (PBS containing 5% BSA). The samples were analyzed using a flow cytometer (FACScanto II, BD).

Western blot analysis

Isolated mouse neutrophils were stimulated with 100 μ g/ml of Scolopendrasin III or Scolopendrasin V for 0 min, 2 min, 5 min, 10 min, and 30 min. Proteins were separated by SDS-PAGE and transferred onto nitrocellulose membranes. The membranes were incubated with specific antibodies (anti-phospho-ERK, anti-phosphor-Akt, or anti-actin), and antigen-antibody complexes were visualized after the membrane was incubated with goat anti-rabbit IgG antibodies combined with horseradish peroxidase. Protein levels were detected using an enhanced chemiluminescence assay as previously described (33).

Generation of bone marrow-derived macrophages

Mouse bone marrow-derived macrophages were generated as previously described (10). Briefly, the bone marrow cells were isolated by flushing the femurs and tibias of wild-type C57BL/6 mice with ice-cold PBS. Bone marrow progenitor cells were cultured in 10 % FBS containing α -MEM under standard incubator conditions for 1 day. After removing the

adherent cells, 10 % FBS containing α -MEM with 30 ng/ml M-CSF was added, and the cells were cultured for 3 days.

***In vivo* neutrophil migration assay**

C57/BL6 mice were purchased from Orient Bio Inc. (Seongnam, Korea). FPR1-deficient mice were kindly provided by Dr. P. Murphy (NIAID, National Institutes of Health, Bethesda, MD, USA) (34). Vehicle or the two AMPs (15mg/kg) were administrated into the peritoneal cavity of WT C57/BL6 mice or FPR1-deficient mice for 2 h. Peritoneal fluid was collected, and then cells were stained with anti-Ly6G or anti-CD11b antibody. Neutrophils that were recruited into the peritoneal fluid were analyzed by flow cytometry.

Data analysis

Results are expressed as mean \pm S.E. The Student's *t*-test was used to compare individual treatments with their respective control values. Statistical significance was set at $p < 0.05$.

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FIGURE LEGENDS

Figure 1. Two novel AMPs selectively stimulate neutrophil chemotaxis via a PTX-sensitive G protein. (A) Alignment of amino acid sequences of three scolopendrasin AMPs. (B) Mouse neutrophils were applied to the upper well of a multiwell chamber

containing several concentrations (0 µg/ml, 1 µg/ml, 5 µg/ml, 10 µg/ml, 50 µg/ml, and 100 µg/ml) of the two AMPs or 1 µM of WKYMVm for 90 min. (C) Mouse neutrophils were

incubated in the absence or presence of 1 µg/ml PTX for 4 h, and were applied to the upper well of the multiwell chamber containing 100 µg/ml of the two AMPs or 1 µM of WKYMVm for 90 min. The number of migrated cells was determined by counting under a

light microscope (B,C). (D) Mouse neutrophils were stimulated with several concentrations (0 µg/ml, 10 µg/ml, 50 µg/ml, and 100 µg/ml) of the two AMPs or 1 µM of WKYMVm and superoxide generated was measured using cytochrome c reduction assay. (E) Several

concentrations (0 µg/ml, 10 µg/ml, 50 µg/ml, and 100 µg/ml) of the two AMPs or 1 µM of WKYMVm were treated to mouse neutrophils for 30 min. The peptide-induced secretion of β-hexosaminidase was determined. Data are presented as means ± S.E. (n=2 for B, C n=3

for D, E). * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ compared to the vehicle-treated control; ### $p < 0.001$ compared to the -PTX control. Data in the panels are representative of at

least three independent experiments performed in duplicate or triplicate.

Figure 2. The two AMPs stimulate ERK and Akt activity in mouse neutrophils. (A)

Fura-2 loaded mouse neutrophils were stimulated with scolopendrasin III (100 µg/ml), scolopendrasin V (100 µg/ml), or 1 µM WKYMVm, and intracellular calcium levels were determined fluorometrically using spectrofluorophotometer. The peak levels of intracellular

calcium were recorded. (B) Mouse neutrophils were stimulated with 100 $\mu\text{g/ml}$ of Scolopendrasin III (left) or Scolopendrasin V (right) for several lengths of time (0 min, 2 min, 5 min, 10 min, 30 min). Total cell lysates were separated SDS-PAGE, and the levels of p-ERK and p-Akt were measured using Western blot analysis. Data are representative of three independent experiments (A, B). (C) Mouse neutrophils were incubated in the absence or presence of PD98059 (50 μM) for 60 min, LY294002 (50 μM) for 15 min, or MK-2206 (2 μM) for 20 min, and were applied to the upper well of a multiwell chamber containing 100 $\mu\text{g/ml}$ of Scolopendrasin III (left) or Scolopendrasin V (right) for 90 min. The number of migrated cells was determined by counting under a light microscope. Data are presented as means \pm S.E. (n=2). Data in the panels are representative of three independent experiments performed in duplicate (C).

Figure 3. Neutrophil chemotaxis stimulated by the two AMPs is mediated by FPR1. (A)

Mouse neutrophils were incubated in the absence or presence of CsH (10 μM) for 30 min, and were applied to the upper well of a multiwell chamber containing 100 $\mu\text{g/ml}$ of the two AMPs or 1 μM of fMLF for 90 min. (B) Cell surface expression of FPR1 was determined by flow cytometric analysis (red, isotype control; blue, anti-FPR1). The results are representative of three independent experiments. (C) Vector- or FPR1-expressing RBL-2H3 cells were applied to the upper well of a multiwell chamber containing several concentrations (0 $\mu\text{g/ml}$, 10 $\mu\text{g/ml}$, 50 $\mu\text{g/ml}$, and 100 $\mu\text{g/ml}$) of the two AMPs or 1 μM of fMLF. (D) Vector- or FPR2-expressing RBL-2H3 cells were applied to the upper well of a multiwell chamber containing several concentrations (0 $\mu\text{g/ml}$, 50 $\mu\text{g/ml}$, and 100 $\mu\text{g/ml}$) of

the two AMPs or 1 μ M of MMK-1. (E) Mouse bone marrow-derived macrophages were applied to the upper well of a multiwell chamber containing several concentrations (0 μ g/ml, 50 μ g/ml, and 100 μ g/ml) of the two AMPs or 1 μ M of WKYMVm for 2 h. The number of migrated cells was determined by counting under a light microscope (A, C, D, E). Data are presented as means \pm S.E. (n=2). * p < 0.05, ** p < 0.01, *** p < 0.001 compared to vehicle-treated (A, E) or vector/RBL2H3 cells (C, D); ### p < 0.001 compared to the AMP alone control (A). Data in the panels are representative of at least three independent experiments performed in duplicate (A, C, D, E).

Figure 4. The two AMPs stimulate neutrophil migration *in vivo* via FPR1. (A) Vehicle, (B) scolopendrasin III (15 mg/kg), or (C) scolopendrasin V (15 mg/kg) were administrated into the peritoneal cavity of WT or FPR1-deficient mice for 2 h. Peritoneal fluid was harvested and recruited neutrophils were analyzed using FACS. Data in the panels are representative of at least three independent experiments. (D) Relative percentages of CD11b⁺ Ly6G⁺ cells from peritoneal fluid were quantified. Data are presented as means \pm S.E. (n=3~6). *** p < 0.001 compared to vehicle-treated; ## p < 0.01 compared to WT control.

Figure 1

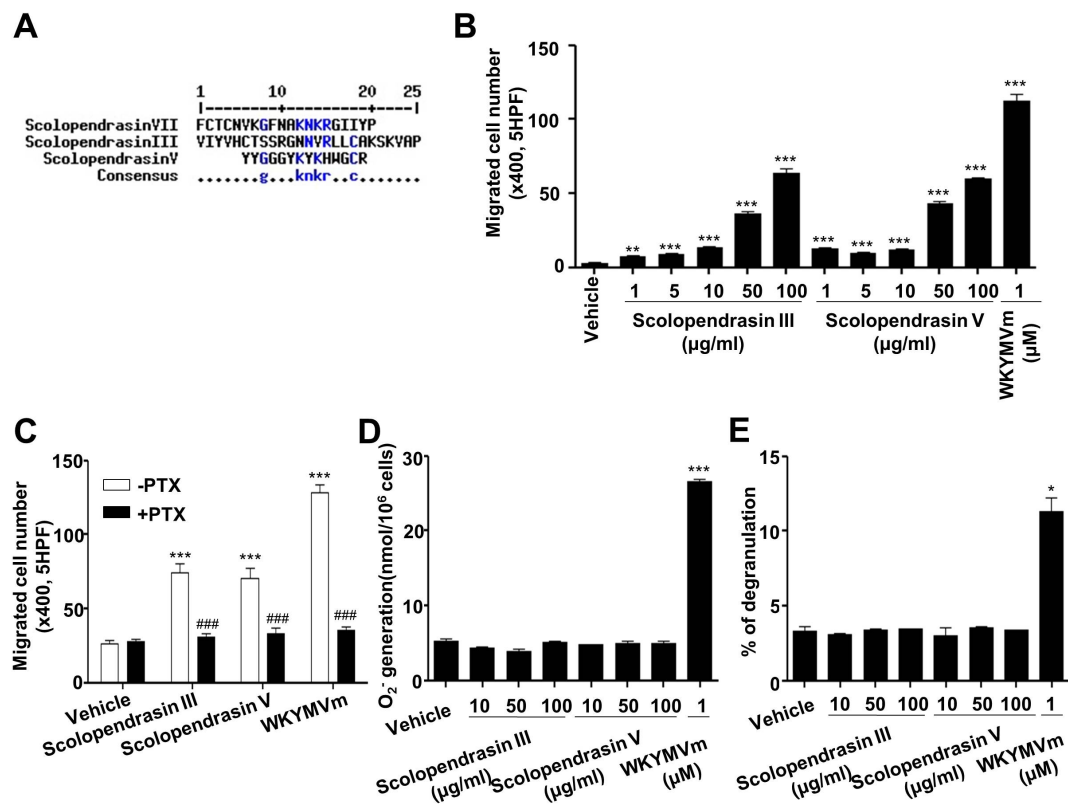


Fig. 1 Figure 1

Figure 2

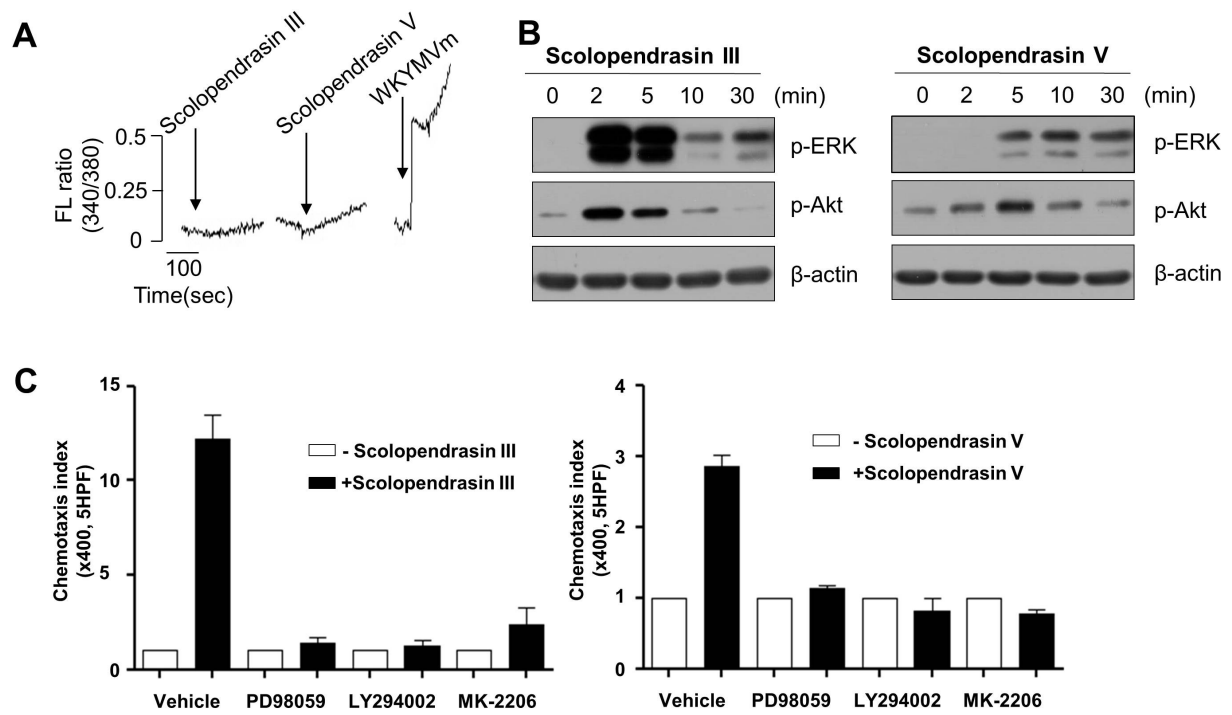


Fig. 2 Figure 2

Figure 3

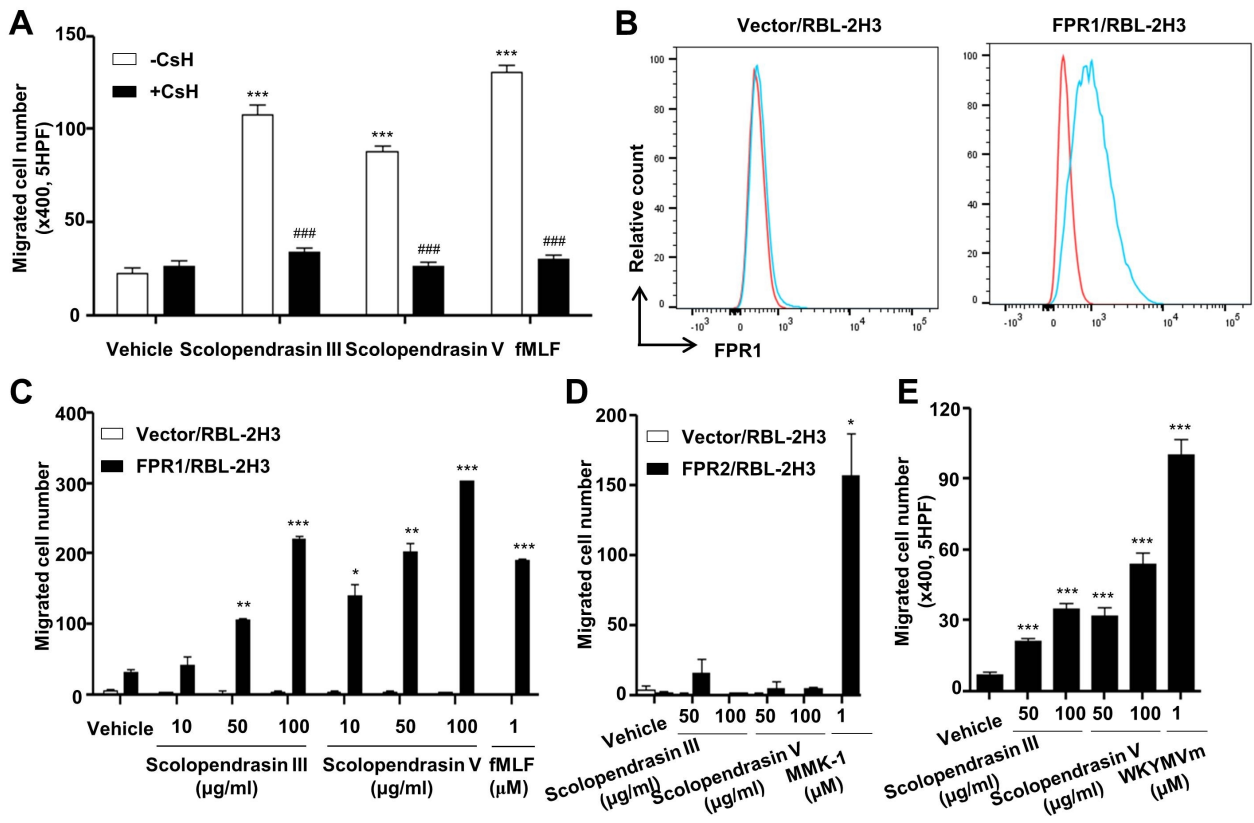


Fig. 3 Figure 3

Figure 4

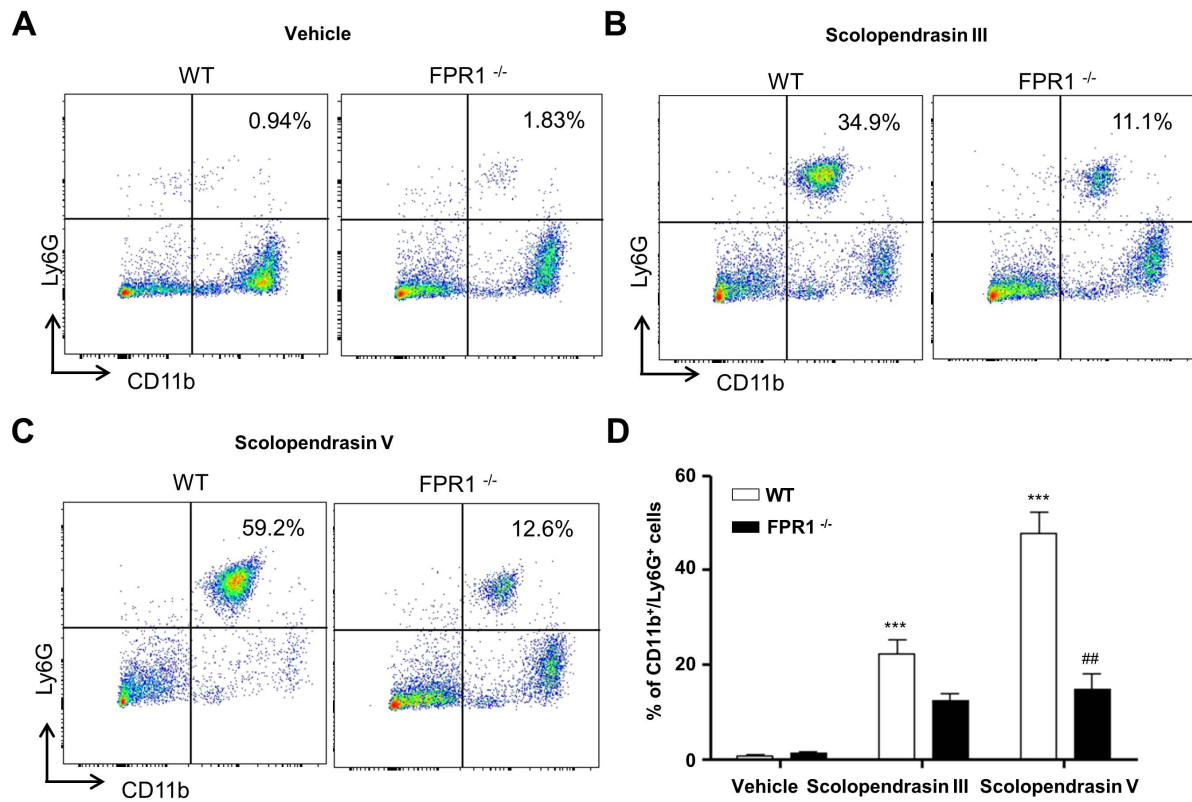


Fig. 4 Figure 4