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**Title:** Function of gaseous hydrogen sulfide in liver fibrosis

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**ABSTRACT**

Over the past few years, hydrogen sulfide ( $H_2S$ ) has been shown to exert several biological functions in mammalian. The endogenous production of  $H_2S$  is mainly mediated by cystathione  $\beta$ -synthase (CBS), cystathione  $\gamma$ -lyase (CSE) and 3-mercaptopyruvate sulfur transferase (MPST). These enzymes are broadly expressed in liver tissue and regulates liver function by working on a variety of molecular targets. As an important regulator of liver function,  $H_2S$  is critically involved in the pathogenesis of various liver diseases, such as non-alcoholic steatohepatitis (NASH), liver fibrosis and liver cancer. Targeting  $H_2S$ -generating enzymes may be a therapeutic strategy for controlling liver diseases. This review described the function of  $H_2S$  in liver disease and summarized recent characterized role of  $H_2S$  in several cellular process of the liver.

## INTRODUCTION

Hydrogen sulfide ( $\text{H}_2\text{S}$ ), well known as a poisonous gas with an unpleasant odour, is produced primarily during the breakdown of proteins in plants and animals (1-3).  $\text{H}_2\text{S}$  is a signaling molecule that is actively synthesized within tissues and is involved in the regulation of vascular tone (4, 5), neuromodulation (6, 7), cell protection (8-10), inflammation (11, 12), and apoptosis (13, 14). Recently, new data on  $\text{H}_2\text{S}$  metabolism and function in animals and humans have been collected under the influence of various endogenous and exogenous factors, including drugs (15, 16).

The liver is one of the most important organs to produce and remove  $\text{H}_2\text{S}$  (17). Endogenous  $\text{H}_2\text{S}$  is involved in the pathogenesis of many liver diseases and affects processes, such as hepatic lipid and glucose metabolism, oxidative stress, mitochondrial bioenergetics, fibrosis, cirrhosis, hepatoprotection, and deregulation of hepatotoxicity (18, 19). In addition, endogenous or exogenous  $\text{H}_2\text{S}$  may play an important role in the development of liver tumors (20, 21). The synthesis and clearance of  $\text{H}_2\text{S}$  in the liver is mainly governed by hepatocytes (17). It is a major source of extracellular matrix (ECM) in hepatic fibrosis and hepatocellular carcinoma (HCC) (22). This review focuses on the major and alternative  $\text{H}_2\text{S}$  metabolism and its regulation in the liver.

### Understanding of $\text{H}_2\text{S}$ metabolism

$\text{H}_2\text{S}$  is a colorless, flammable gas with a characteristic odor of rotten eggs. It occurs naturally in volcanic gases, natural gas, and some well water, and is also produced when bacteria decompose organic matter in the absence of oxygen (23).  $\text{H}_2\text{S}$  is toxic to humans and can result in death from acute exposure to large amounts of  $\text{H}_2\text{S}$  (>500 ppm) (24).  $\text{H}_2\text{S}$  was considered both a toxic molecule and an environmental hazard until discovered to be endogenously

produced (1). The production of H<sub>2</sub>S by three enzymes like cystathionine  $\beta$ -synthetase (CBS), cystathionine  $\gamma$ -lyase (CSE) and 3-mercaptopyruvate sulfur transferase (MPST) (25-28) has been widely studied (Fig. 1). Endogenous H<sub>2</sub>S is produced by enzymatic activity and is also released from intracellular sulfur stores (29). In most organ, CBS and CSE are mainly responsible for H<sub>2</sub>S production (29). They manage individually from L-cysteine to produce H<sub>2</sub>S, L-serine and ammonium (30). Although found throughout the body, the discovery of CBS in the brain has led to consensus that it is a major H<sub>2</sub>S-generating enzyme that affects nerve signaling (31). However, CBS has been identified in tissues throughout the body and is thought to regulate overall H<sub>2</sub>S production (32). Located primarily in mitochondria, MPST enzymatically generates H<sub>2</sub>S from  $\alpha$ -ketoglutarate and L-cysteine through metabolic interactions with cysteine aminotransferase (33). CBS, CSE and MPST are mainly expressed in the liver and kidney (34). CBS and CSE metabolize cysteine and/or homocysteine to release H<sub>2</sub>S (35), while MPST metabolizes cysteine and 3-mercaptopyruvic acid (3-MP) produced by the action of cysteine aminotransferase (CAT) on  $\alpha$ -ketoglutaric acid (36, 37). MPST requires a cofactor to decrease the persulfate intermediate formed between the MPST cysteine residue and the sulfide provided by 3-MP (36). Recent data have found that thioredoxin and dihydrolipoic acid (DHLA) are endogenous reduction cofactors which promote H<sub>2</sub>S release from MPST (38).

H<sub>2</sub>S is an endogenous signaling molecule in mammals (39). Accumulating evidence suggests that H<sub>2</sub>S plays an important role in liver physiology and pathophysiology (40-42). Dysregulation of endogenous H<sub>2</sub>S is associated with symptoms of diabetes and cirrhosis (43, 44). Blood levels of H<sub>2</sub>S in patients with type 2 diabetes mellitus are lower than in controls (45). Application of H<sub>2</sub>S also shows effects on mitochondrial function, antioxidant stress, apoptosis, inflammation, angiogenesis, and blood pressure (46).

### Function of H<sub>2</sub>S in the liver

The liver plays an important role in mammalian physiology with respect to energy homeostasis (47). Besides, the liver is also a major detoxification tissue, and can metabolize and neutralize harmful substances, drugs, environmental toxins, and endotoxins (48, 49). Endogenous formation of H<sub>2</sub>S is impaired in non-alcoholic steatohepatitis (NASH) mice, and H<sub>2</sub>S treatment can prevent NASH in mice, perhaps by reducing oxidative stress and suppressing inflammation (40). Administration of sodium hydrogen sulfide (NaHS) as a H<sub>2</sub>S donor in rodents protects against ischemic reperfusion, acetaminophen or carbon tetrachloride (CCl<sub>4</sub>)-induced liver damage (50).

The liver is uniquely positioned to be exposed to high levels of H<sub>2</sub>S; however, how the liver responds to elevated hydrogen sulfide levels is unclear. Liver H<sub>2</sub>S levels were previously reported within the low nanomolar to middle micromolar range (17 nM ~ 144 μM) (51). Reactive oxygen species (ROS), a by-product of normal aerobic cell metabolism, are important signaling molecules in many cell functions such as immune response, apoptosis and cell survival (52-54). Recent studies have shown that treatment with relatively low concentrations of H<sub>2</sub>S donors such as NaHS, Na<sub>2</sub>S or GYY4137 (50 mg/kg) may decrease ROS levels and cytochrome P450 2E1 activity and increase glutathione levels and antioxidant enzymes (50, 55). These results indicate that relatively low levels of H<sub>2</sub>S can protect against oxidative stress in the liver. Mitochondria is bilayer organelles whose shape supports them function in many cellular processes (56). The main role of mitochondria is to regulate the production of energetic molecules like adenosine triphosphate (57). During the metabolism of glucose, lipids, and proteins in the liver (58), 3-MP, the substrate for the MPST, stimulates mitochondrial H<sub>2</sub>S production and enhances liver mitochondrial electron transport at low concentrations (59, 60).

In addition, low levels of H<sub>2</sub>S induces a significant increase in hepatic mitochondrial function (61). Moreover, H<sub>2</sub>S acts on mitochondrial proteins via a posttranslational modification designated as sulfhydration or persulfidation (62, 63). Sulfhydration of the ATP Synthase F1 Subunit Alpha (ATP5A1) at Cys244 and 294 was reported to increase its activity (64). Sulfhydration of ATP5A1 was upregulated in response to burn injury and decreased in mice lacking CSE implicating a role for CSE-derived H<sub>2</sub>S in the process (64). These results indicate that endogenous H<sub>2</sub>S regulates physiologically in mitochondrial electron transport.

The liver is important for the maintenance of blood glucose homeostasis by the uptake of glucose in the postprandial state and its conversion to triglycerides and glycogen, and the production of glucose in the post-absorption state by gluconeogenesis and glycogenolysis (65, 66). Deficiencies in the mechanism by which insulin and glucose regulate glycogen metabolism in the liver disrupt blood glucose homeostasis, leading to metabolic disorders such as diabetes and glycogen storage (67, 68). CSE activity has been shown to be low in the liver of type 1 diabetic rats and in peripheral blood mononuclear cells of type 1 diabetic patients, indicating that H<sub>2</sub>S is involved in glucose regulation (69). Recent studies have shown that CSE knockout mice have a reduced rate of glycolysis. This can be reversed with NaHS management (70, 71). NAFLD is caused by the accumulation of lipids in the liver and may increase the risk of hepatocellular carcinoma and end-stage liver disease (72, 73). Many risk factors, such as diabetes, obesity, hyperlipidemia, and certain drug regimens are associated with the development of NAFLD (74). H<sub>2</sub>S has been shown to alleviate development of fatty liver in obese mice through its antioxidant capacity and promotion of lipid metabolism (40, 75). In a recent study, in NAFLD mouse model, the activation of sterol regulatory element binding protein-1c directly upregulates mir-216a transcription, which reduces CTH-H<sub>2</sub>S signaling and ULK1-stimulated autophagy, indicating that loss of sterol regulatory element binding protein-

1c prevents the development of hepatic steatosis through activation of H<sub>2</sub>S-mediated autophagy flux in a high fat diets-induced NAFLD model (76, 77). Recent study has shown that administration of NaHS reduces the accumulation of lipids such as total cholesterol and triglycerides through down-regulation of fatty acid synthase and up-regulation of carnitine palmitoyl transferase-1 in the liver of high-fat diet (HFD)-induced obese mice (40). Collectively, H<sub>2</sub>S may alleviate liver cell damage in various ways in the pathogenesis of liver disease (Fig 1).

### **Role of H<sub>2</sub>S in liver fibrosis**

Several studies have been reported on the use of H<sub>2</sub>S in hypoxic injury (78, 79), most of which show beneficial effects of H<sub>2</sub>S treatment in models of cardiac arrest (80), lung (81), intestinal (82), renal (83), and cardiac ischemia (84). Fibrogenesis formation in chronic liver disease can disrupt liver functional units and blood flow, leading to cirrhosis of the liver and even life-threatening clinical outcomes (85, 86). In the pathological process of hepatic fibrosis, it is widely known that activated hepatic stellate cells (HSC) are fundamental to the overproduction of ECM in the stroma (87). Recent evidence suggests that inactivation of HSC is an essential mechanism by which H<sub>2</sub>S inhibits liver fibrosis (88). However, current report shows that the generation of H<sub>2</sub>S is increased during HSC activation, and that exogenous H<sub>2</sub>S promotes HSC proliferation and induces the expression of HSC fibrosis makers (89). Furthermore, conflicting results have also been reported depending on the concentration or type of H<sub>2</sub>S donor used. Based on the H<sub>2</sub>S release rate, H<sub>2</sub>S release donors are classified as either fast (NaHS; Na<sub>2</sub>S) or slow (GYY4137; ADT-OH) release donors, often giving contrasting results (90, 91). For example, some studies have reported pro-inflammatory and anti-apoptotic properties of H<sub>2</sub>S, and shown that H<sub>2</sub>S increases mitochondrial bioenergetics and promotes cell proliferation (64,



92, 93). Therefore, there is still a large gap in our understanding of the actual impact of H<sub>2</sub>S on HSC and liver fibrosis.

The CCl<sub>4</sub>-induced hepatic fibrosis model tends to suppress protein expression of both CSE and H<sub>2</sub>S content (94). Suggestion for a protective function for H<sub>2</sub>S in liver fibrosis is supported by the understanding that CBS deficiency accelerates fibrosis associated with hepatic steatosis (95). Similarly, gene knockout of CSE exacerbates liver fibrosis by triggering an inflammatory response and decreasing H<sub>2</sub>S production, indicating a potential role of the H<sub>2</sub>S system in liver fibrosis (96). Supplementation of NaHS ameliorates hepatic fibrosis in CCl<sub>4</sub>-treated mice (50). Likewise, CCl<sub>4</sub>-treated mice, GYY4137, increased nuclear factor erythroid 2–related factor 2 signaling pathway, improved liver function, reduced liver fibrosis, decreased hepatic oxidative stress (97). Exercise significantly enhances H<sub>2</sub>S level and increases levels of CBS, CSE and MPST in HFD-fed mice (98).

H<sub>2</sub>S reduces the intracellular redox environment and reduces damage from oxidative stress (99). Given the important role of oxidative stress in the development of fibrosis, it is reasonable to suspect that the endogenous H<sub>2</sub>S-producing enzyme pathway suppresses the development of fibrosis by its antioxidant activity (100). Extrinsic H<sub>2</sub>S inhibits Fe-NTA-induced elevated intracellular ROS levels and HSC cell proliferation (94), weakens CCl<sub>4</sub>-induced increase of liver MDA levels, reduces liver GSH levels, and collagen in liver tissue. It is associated with inhibition of phosphorylated p38 mitogen-activated protein kinase and activation of the phospho-AKT signaling pathway (101).

Inflammation has been reported to be in the early stages of the onset of fibrosis, causing cell apoptosis, fibroblast proliferation, and ECM deposition, ultimately leading to irreversible fibrous damage (102). Treatment with H<sub>2</sub>S significantly reduces the infiltration of inflammatory cells, inducible nitrogen monoxide synthase, tumor necrosis factor- $\alpha$ , It down-

regulates pro-inflammatory cytokines such as interleukin (IL)-6, and inhibits IL-8, and the progression of fibrosis (18, 103-105). Although CCl<sub>4</sub>-induced liver cirrhosis rats showed significantly higher levels of serum inflammation-inducing cytokines. Co-administration of NaHS resulted in a significant reduction in these cytokines, along with the alleviated collagen fibers of the liver (50).

Recent studies have shown that organ fibrosis is associated with a decrease in autophagy (106, 107). Autophagy is involved in a complex regulatory pathway in hepatic fibrosis, and its fibrosis-promoting effect depends on the activation of hepatic stellate cells but has antifibrotic properties through indirect hepatic protection and anti-inflammatory properties (108). Given the important role of autophagy in the pathogenesis of fibrosis and the regulatory function of H<sub>2</sub>S for autophagy and fibrosis, extrinsic or endogenous H<sub>2</sub>S is mediated by targeting by autophagy or autophagy-related signaling pathways (21, 109). It is rational and interesting to assume that it may inhibit the development of fibrosis. Overall, these observations suggest that an endogenous H<sub>2</sub>S system or H<sub>2</sub>S-releasing donor can be developed to treat liver fibrosis via a variety of signaling pathways (Fig. 2).

## CONCLUSION

This review summarizes and describes the recent literature on the role of H<sub>2</sub>S in several liver diseases. Defect in endogenous H<sub>2</sub>S production is associated with NASH and liver fibrosis. And because H<sub>2</sub>S may serve as a double-edged sword in such liver disorder, additional studies need to resolve these discrepancies in the future. In addition, although endogenous H<sub>2</sub>S production or low exogenous H<sub>2</sub>S may lead to the development of liver fibrosis, exposure to large amounts of H<sub>2</sub>S may exhibit anti-fibrosis properties. Therefore, targeting H<sub>2</sub>S-producing enzymes may be a promising strategy for managing liver disorders.

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## **CONFLICTS OF INTEREST**

The authors have no conflicting interests.

## REFERENCES

1. Beauchamp RO, Jr., Bus JS, Popp JA, Boreiko CJ and Andjelkovich DA (1984) A critical review of the literature on hydrogen sulfide toxicity. *Crit Rev Toxicol* 13, 25-97
2. Wang R (2002) Two's company, three's a crowd: can H<sub>2</sub>S be the third endogenous gaseous transmitter? *FASEB J* 16, 1792-1798
3. Aroca A, Gotor C and Romero LC (2018) Hydrogen Sulfide Signaling in Plants: Emerging Roles of Protein Persulfidation. *Front Plant Sci* 9, 1369
4. Dombkowski RA, Russell MJ and Olson KR (2004) Hydrogen sulfide as an endogenous regulator of vascular smooth muscle tone in trout. *Am J Physiol Regul Integr Comp Physiol* 286, R678-685
5. Wojcicka G, Jamroz-Wisniewska A, Atanasova P, Chaldakov GN, Chylinska-Kula B and Beltowski J (2011) Differential effects of statins on endogenous H<sub>2</sub>S formation in perivascular adipose tissue. *Pharmacol Res* 63, 68-76
6. Abe K and Kimura H (1996) The possible role of hydrogen sulfide as an endogenous neuromodulator. *J Neurosci* 16, 1066-1071
7. Kimura H (2002) Hydrogen sulfide as a neuromodulator. *Mol Neurobiol* 26, 13-19
8. Li L, Rose P and Moore PK (2011) Hydrogen sulfide and cell signaling. *Annu Rev Pharmacol Toxicol* 51, 169-187
9. Xiao Q, Ying J, Xiang L and Zhang C (2018) The biologic effect of hydrogen sulfide and its function in various diseases. *Medicine (Baltimore)* 97, e13065
10. Han Y, Shang Q, Yao J and Ji Y (2019) Hydrogen sulfide: a gaseous signaling molecule modulates tissue homeostasis: implications in ophthalmic diseases. *Cell Death Dis* 10, 293
11. Whiteman M and Winyard PG (2011) Hydrogen sulfide and inflammation: the good, the bad, the ugly and the promising. *Expert Rev Clin Pharmacol* 4, 13-32
12. Bhatia M and Gaddam RR (2021) Hydrogen Sulfide in Inflammation: A Novel

Mediator and Therapeutic Target. *Antioxid Redox Signal* 34, 1368-1377

13. Ryazantseva NV, Novitsky VV, Starikova EG, Kleptsova LA, Jakushina VD and Kaigorodova EV (2011) Role of hydrogen sulfide in the regulation of cell apoptosis. *Bull Exp Biol Med* 151, 702-704
14. Li X, Chen M, Shi Q, Zhang H and Xu S (2020) Hydrogen sulfide exposure induces apoptosis and necroptosis through lncRNA3037/miR-15a/BCL2-A20 signaling in broiler trachea. *Sci Total Environ* 699, 134296
15. Zaichko NV, Melnik AV, Yoltukhivskyy MM, Olhovskiy AS and Palamarchuk IV (2014) Hydrogen sulfide: metabolism, biological and medical role. *Ukr Biochem J* 86, 5-25
16. Arif MS, Yasmeen T, Abbas Z et al (2020) Role of Exogenous and Endogenous Hydrogen Sulfide (H<sub>2</sub>S) on Functional Traits of Plants Under Heavy Metal Stresses: A Recent Perspective. *Front Plant Sci* 11, 545453
17. Norris EJ, Culbertson CR, Narasimhan S and Clemens MG (2011) The liver as a central regulator of hydrogen sulfide. *Shock* 36, 242-250
18. Wu DD, Wang DY, Li HM, Guo JC, Duan SF and Ji XY (2019) Hydrogen Sulfide as a Novel Regulatory Factor in Liver Health and Disease. *Oxid Med Cell Longev* 2019, 3831713
19. Sun HJ, Wu ZY, Nie XW, Wang XY and Bian JS (2021) Implications of hydrogen sulfide in liver pathophysiology: Mechanistic insights and therapeutic potential. *J Adv Res* 27, 127-135
20. Hellmich MR and Szabo C (2015) Hydrogen Sulfide and Cancer. *Handb Exp Pharmacol* 230, 233-241
21. Wang SS, Chen YH, Chen N et al (2017) Hydrogen sulfide promotes autophagy of hepatocellular carcinoma cells through the PI3K/Akt/mTOR signaling pathway. *Cell Death Dis* 8, e2688
22. Filliol A and Schwabe RF (2019) Contributions of Fibroblasts, Extracellular Matrix, Stiffness, and Mechanosensing to Hepatocarcinogenesis. *Semin Liver Dis* 39, 315-333

23. Malone Rubright SL, Pearce LL and Peterson J (2017) Environmental toxicology of hydrogen sulfide. *Nitric Oxide* 71, 1-13
24. Doujaiji B and Al-Tawfiq JA (2010) Hydrogen sulfide exposure in an adult male. *Ann Saudi Med* 30, 76-80
25. Ahmad A, Gero D, Olah G and Szabo C (2016) Effect of endotoxemia in mice genetically deficient in cystathionine-gamma-lyase, cystathionine-beta-synthase or 3-mercaptopyruvate sulfurtransferase. *Int J Mol Med* 38, 1683-1692
26. Tao B, Wang R, Sun C and Zhu Y (2017) 3-Mercaptopyruvate Sulfurtransferase, Not Cystathionine beta-Synthase Nor Cystathionine gamma-Lyase, Mediates Hypoxia-Induced Migration of Vascular Endothelial Cells. *Front Pharmacol* 8, 657
27. Ahmad A, Druzhyna N and Szabo C (2019) Effect of 3-mercaptopyruvate Sulfurtransferase Deficiency on the Development of Multiorgan Failure, Inflammation, and Wound Healing in Mice Subjected to Burn Injury. *J Burn Care Res* 40, 148-156
28. Augsburger F and Szabo C (2020) Potential role of the 3-mercaptopyruvate sulfurtransferase (3-MST)-hydrogen sulfide (H<sub>2</sub>S) pathway in cancer cells. *Pharmacol Res* 154, 104083
29. Cao X, Ding L, Xie ZZ et al (2019) A Review of Hydrogen Sulfide Synthesis, Metabolism, and Measurement: Is Modulation of Hydrogen Sulfide a Novel Therapeutic for Cancer? *Antioxid Redox Signal* 31, 1-38
30. Shibuya N, Koike S, Tanaka M et al (2013) A novel pathway for the production of hydrogen sulfide from D-cysteine in mammalian cells. *Nat Commun* 4, 1366
31. Polhemus DJ and Lefer DJ (2014) Emergence of hydrogen sulfide as an endogenous gaseous signaling molecule in cardiovascular disease. *Circ Res* 114, 730-737
32. Murphy B, Bhattacharya R and Mukherjee P (2019) Hydrogen sulfide signaling in mitochondria and disease. *FASEB J* 33, 13098-13125
33. Pedre B and Dick TP (2021) 3-Mercaptopyruvate sulfurtransferase: an enzyme at the crossroads of sulfane sulfur trafficking. *Biol Chem* 402, 223-237

34. Stipanuk MH and Beck PW (1982) Characterization of the enzymic capacity for cysteine desulphhydration in liver and kidney of the rat. *Biochem J* 206, 267-277
35. Nandi SS and Mishra PK (2017) H<sub>2</sub>S and homocysteine control a novel feedback regulation of cystathionine beta synthase and cystathionine gamma lyase in cardiomyocytes. *Sci Rep* 7, 3639
36. Cooper AJ (1983) Biochemistry of sulfur-containing amino acids. *Annu Rev Biochem* 52, 187-222
37. Hipolito A, Nunes SC, Vicente JB and Serpa J (2020) Cysteine Aminotransferase (CAT): A Pivotal Sponsor in Metabolic Remodeling and an Ally of 3-Mercaptopyruvate Sulfurtransferase (MST) in Cancer. *Molecules* 25
38. Mikami Y and Kimura H (2012) A mechanism of retinal protection from light-induced degeneration by hydrogen sulfide. *Commun Integr Biol* 5, 169-171
39. Guo W, Kan JT, Cheng ZY et al (2012) Hydrogen sulfide as an endogenous modulator in mitochondria and mitochondria dysfunction. *Oxid Med Cell Longev* 2012, 878052
40. Wu D, Zheng N, Qi K et al (2015) Exogenous hydrogen sulfide mitigates the fatty liver in obese mice through improving lipid metabolism and antioxidant potential. *Med Gas Res* 5, 1
41. Wu D, Zhong P, Wang Y et al (2020) Hydrogen Sulfide Attenuates High-Fat Diet-Induced Non-Alcoholic Fatty Liver Disease by Inhibiting Apoptosis and Promoting Autophagy via Reactive Oxygen Species/Phosphatidylinositol 3-Kinase/AKT/Mammalian Target of Rapamycin Signaling Pathway. *Front Pharmacol* 11, 585860
42. Comas F and Moreno-Navarrete JM (2021) The Impact of H<sub>2</sub>S on Obesity-Associated Metabolic Disturbances. *Antioxidants (Basel)* 10
43. Wang P and Wu L (2018) Hydrogen sulfide and nonalcoholic fatty liver disease. *Hepatobiliary Surg Nutr* 7, 122-124
44. Carter RN, Gibbins MTG, Barrios-Llerena ME et al (2021) The hepatic compensatory

- response to elevated systemic sulfide promotes diabetes. *Cell Rep* 37, 109958
45. Pineiro-Ramil M, Burguera EF, Hermida-Gomez T et al (2022) Reduced Levels of H<sub>2</sub>S in Diabetes-Associated Osteoarthritis Are Linked to Hyperglycaemia, Nrf-2/HO-1 Signalling Downregulation and Chondrocyte Dysfunction. *Antioxidants (Basel)* 11
  46. Gorini F, Del Turco S, Sabatino L, Gaggini M and Vassalle C (2021) H<sub>2</sub>S as a Bridge Linking Inflammation, Oxidative Stress and Endothelial Biology: A Possible Defense in the Fight against SARS-CoV-2 Infection? *Biomedicines* 9
  47. Jensen-Cody SO and Potthoff MJ (2021) Hepatokines and metabolism: Deciphering communication from the liver. *Mol Metab* 44, 101138
  48. Grant DM (1991) Detoxification pathways in the liver. *J Inherit Metab Dis* 14, 421-430
  49. Melaram R (2021) Environmental Risk Factors Implicated in Liver Disease: A Mini-Review. *Front Public Health* 9, 683719
  50. Tan G, Pan S, Li J et al (2011) Hydrogen sulfide attenuates carbon tetrachloride-induced hepatotoxicity, liver cirrhosis and portal hypertension in rats. *PLoS One* 6, e25943
  51. Mateus I and Prip-Buus C (2022) Hydrogen sulphide in liver glucose/lipid metabolism and non-alcoholic fatty liver disease. *Eur J Clin Invest* 52, e13680
  52. Previte DM, O'Connor EC, Novak EA, Martins CP, Mollen KP and Piganelli JD (2017) Reactive oxygen species are required for driving efficient and sustained aerobic glycolysis during CD4<sup>+</sup> T cell activation. *PLoS One* 12, e0175549
  53. Snezhkina AV, Kudryavtseva AV, Kardymon OL et al (2019) ROS Generation and Antioxidant Defense Systems in Normal and Malignant Cells. *Oxid Med Cell Longev* 2019, 6175804
  54. Peng HY, Lucavs J, Ballard D et al (2021) Metabolic Reprogramming and Reactive Oxygen Species in T Cell Immunity. *Front Immunol* 12, 652687
  55. Li L, Salto-Tellez M, Tan CH, Whiteman M and Moore PK (2009) GYY4137, a novel hydrogen sulfide-releasing molecule, protects against endotoxic shock in the rat. *Free*



Radic Biol Med 47, 103-113

56. Xia M, Zhang Y, Jin K, Lu Z, Zeng Z and Xiong W (2019) Communication between mitochondria and other organelles: a brand-new perspective on mitochondria in cancer. *Cell Biosci* 9, 27
57. Brand MD, Orr AL, Perevoshchikova IV and Quinlan CL (2013) The role of mitochondrial function and cellular bioenergetics in ageing and disease. *Br J Dermatol* 169 Suppl 2, 1-8
58. Degli Esposti D, Hamelin J, Bosselut N et al (2012) Mitochondrial roles and cytoprotection in chronic liver injury. *Biochem Res Int* 2012, 387626
59. Modis K, Coletta C, Erdelyi K, Papapetropoulos A and Szabo C (2013) Intramitochondrial hydrogen sulfide production by 3-mercaptopyruvate sulfurtransferase maintains mitochondrial electron flow and supports cellular bioenergetics. *FASEB J* 27, 601-611
60. Jia J, Wang Z, Zhang M et al (2020) SQR mediates therapeutic effects of H<sub>2</sub>S by targeting mitochondrial electron transport to induce mitochondrial uncoupling. *Sci Adv* 6, eaaz5752
61. Paul BD, Snyder SH and Kashfi K (2021) Effects of hydrogen sulfide on mitochondrial function and cellular bioenergetics. *Redox Biol* 38, 101772
62. Shimizu Y, Polavarapu R, Eskla KL et al (2018) Hydrogen sulfide regulates cardiac mitochondrial biogenesis via the activation of AMPK. *J Mol Cell Cardiol* 116, 29-40
63. Mustafa AK, Gadalla MM, Sen N et al (2009) H<sub>2</sub>S signals through protein S-sulfhydration. *Sci Signal* 2, ra72
64. Modis K, Ju Y, Ahmad A et al (2016) S-Sulfhydration of ATP synthase by hydrogen sulfide stimulates mitochondrial bioenergetics. *Pharmacol Res* 113, 116-124
65. Nuttall FQ, Ngo A and Gannon MC (2008) Regulation of hepatic glucose production and the role of gluconeogenesis in humans: is the rate of gluconeogenesis constant? *Diabetes Metab Res Rev* 24, 438-458

66. Cruz-Pineda WD, Parra-Rojas I, Rodriguez-Ruiz HA, Illades-Aguilar B, Matia-Garcia I and Garibay-Cerdenares OL (2022) The regulatory role of insulin in energy metabolism and leukocyte functions. *J Leukoc Biol* 111, 197-208
67. Han HS, Kang G, Kim JS, Choi BH and Koo SH (2016) Regulation of glucose metabolism from a liver-centric perspective. *Exp Mol Med* 48, e218
68. Irimia JM, Meyer CM, Segvich DM et al (2017) Lack of liver glycogen causes hepatic insulin resistance and steatosis in mice. *J Biol Chem* 292, 10455-10464
69. Manna P, Gungor N, McVie R and Jain SK (2014) Decreased cystathionine-gamma-lyase (CSE) activity in livers of type 1 diabetic rats and peripheral blood mononuclear cells (PBMC) of type 1 diabetic patients. *J Biol Chem* 289, 11767-11778
70. Untereiner AA, Wang R, Ju Y and Wu L (2016) Decreased Gluconeogenesis in the Absence of Cystathionine Gamma-Lyase and the Underlying Mechanisms. *Antioxid Redox Signal* 24, 129-140
71. Li N, Wang MJ, Jin S et al (2016) The H<sub>2</sub>S Donor NaHS Changes the Expression Pattern of H<sub>2</sub>S-Producing Enzymes after Myocardial Infarction. *Oxid Med Cell Longev* 2016, 6492469
72. Dhamija E, Paul SB and Kedia S (2019) Non-alcoholic fatty liver disease associated with hepatocellular carcinoma: An increasing concern. *Indian J Med Res* 149, 9-17
73. Rada P, Gonzalez-Rodriguez A, Garcia-Monzon C and Valverde AM (2020) Understanding lipotoxicity in NAFLD pathogenesis: is CD36 a key driver? *Cell Death Dis* 11, 802
74. Raman M and Allard J (2006) Non alcoholic fatty liver disease: a clinical approach and review. *Can J Gastroenterol* 20, 345-349
75. Li M, Xu C, Shi J et al (2018) Fatty acids promote fatty liver disease via the dysregulation of 3-mercaptopyruvate sulfurtransferase/hydrogen sulfide pathway. *Gut* 67, 2169-2180
76. Nguyen TTP, Kim DY, Lee YG et al (2021) SREBP-1c impairs ULK1 sulphydration-

mediated autophagic flux to promote hepatic steatosis in high-fat-diet-fed mice. *Mol Cell* 81, 3820-3832 e3827

77. Nguyen TTP, Kim DY, Im SS and Jeon TI (2021) Impairment of ULK1 sulfhydration-mediated lipophagy by SREBF1/SREBP-1c in hepatic steatosis. *Autophagy* 17, 4489-4490
78. Lan A, Liao X, Mo L et al (2011) Hydrogen sulfide protects against chemical hypoxia-induced injury by inhibiting ROS-activated ERK1/2 and p38MAPK signaling pathways in PC12 cells. *PLoS One* 6, e25921
79. Hine C, Harputlugil E, Zhang Y et al (2015) Endogenous hydrogen sulfide production is essential for dietary restriction benefits. *Cell* 160, 132-144
80. Minamishima S, Bougaki M, Sips PY et al (2009) Hydrogen sulfide improves survival after cardiac arrest and cardiopulmonary resuscitation via a nitric oxide synthase 3-dependent mechanism in mice. *Circulation* 120, 888-896
81. Jiang T, Yang W, Zhang H, Song Z, Liu T and Lv X (2020) Hydrogen Sulfide Ameliorates Lung Ischemia-Reperfusion Injury Through SIRT1 Signaling Pathway in Type 2 Diabetic Rats. *Front Physiol* 11, 596
82. Liu H, Bai XB, Shi S and Cao YX (2009) Hydrogen sulfide protects from intestinal ischaemia-reperfusion injury in rats. *J Pharm Pharmacol* 61, 207-212
83. Sekijima M, Sahara H, Miki K et al (2017) Hydrogen sulfide prevents renal ischemia-reperfusion injury in CLAWN miniature swine. *J Surg Res* 219, 165-172
84. Elrod JW, Calvert JW, Morrison J et al (2007) Hydrogen sulfide attenuates myocardial ischemia-reperfusion injury by preservation of mitochondrial function. *Proc Natl Acad Sci U S A* 104, 15560-15565
85. Takahashi H, Shigefuku R, Yoshida Y et al (2014) Correlation between hepatic blood flow and liver function in alcoholic liver cirrhosis. *World J Gastroenterol* 20, 17065-17074
86. Iwakiri Y, Shah V and Rockey DC (2014) Vascular pathobiology in chronic liver disease

- and cirrhosis - current status and future directions. *J Hepatol* 61, 912-924
87. Coulouarn C and Clement B (2014) Stellate cells and the development of liver cancer: therapeutic potential of targeting the stroma. *J Hepatol* 60, 1306-1309
  88. Zhang F, Jin H, Wu L et al (2017) Diallyl Trisulfide Suppresses Oxidative Stress-Induced Activation of Hepatic Stellate Cells through Production of Hydrogen Sulfide. *Oxid Med Cell Longev* 2017, 1406726
  89. Damba T, Zhang M, Buist-Homan M, van Goor H, Faber KN and Moshage H (2019) Hydrogen sulfide stimulates activation of hepatic stellate cells through increased cellular bio-energetics. *Nitric Oxide* 92, 26-33
  90. Wedmann R, Bertlein S, Macinkovic I et al (2014) Working with "H<sub>2</sub>S": facts and apparent artifacts. *Nitric Oxide* 41, 85-96
  91. Zheng Y, Ji X, Ji K and Wang B (2015) Hydrogen sulfide prodrugs-a review. *Acta Pharm Sin B* 5, 367-377
  92. Xie X, Dai H, Zhuang B, Chai L, Xie Y and Li Y (2016) Exogenous hydrogen sulfide promotes cell proliferation and differentiation by modulating autophagy in human keratinocytes. *Biochem Biophys Res Commun* 472, 437-443
  93. Phillips CM, Zatarain JR, Nicholls ME et al (2017) Upregulation of Cystathionine-beta-Synthase in Colonic Epithelia Reprograms Metabolism and Promotes Carcinogenesis. *Cancer Res* 77, 5741-5754
  94. Fan HN, Wang HJ, Yang-Dan CR et al (2013) Protective effects of hydrogen sulfide on oxidative stress and fibrosis in hepatic stellate cells. *Mol Med Rep* 7, 247-253
  95. Robert K, Nehme J, Bourdon E et al (2005) Cystathionine beta synthase deficiency promotes oxidative stress, fibrosis, and steatosis in mice liver. *Gastroenterology* 128, 1405-1415
  96. Ci L, Yang X, Gu X et al (2017) Cystathionine gamma-Lyase Deficiency Exacerbates CCl<sub>4</sub>-Induced Acute Hepatitis and Fibrosis in the Mouse Liver. *Antioxid Redox Signal* 27, 133-149

97. Zhao S, Song T, Gu Y et al (2021) Hydrogen Sulfide Alleviates Liver Injury Through the S-Sulhydrated-Kelch-Like ECH-Associated Protein 1/Nuclear Erythroid 2-Related Factor 2/Low-Density Lipoprotein Receptor-Related Protein 1 Pathway. *Hepatology* 73, 282-302
98. Wang B, Zeng J and Gu Q (2017) Exercise restores bioavailability of hydrogen sulfide and promotes autophagy influx in livers of mice fed with high-fat diet. *Can J Physiol Pharmacol* 95, 667-674
99. Scammahorn JJ, Nguyen ITN, Bos EM, Van Goor H and Joles JA (2021) Fighting Oxidative Stress with Sulfur: Hydrogen Sulfide in the Renal and Cardiovascular Systems. *Antioxidants (Basel)* 10
100. Zhang S, Pan C, Zhou F et al (2015) Hydrogen Sulfide as a Potential Therapeutic Target in Fibrosis. *Oxid Med Cell Longev* 2015, 593407
101. Fan HN, Wang HJ, Ren L et al (2013) Decreased expression of p38 MAPK mediates protective effects of hydrogen sulfide on hepatic fibrosis. *Eur Rev Med Pharmacol Sci* 17, 644-652
102. Wynn TA and Ramalingam TR (2012) Mechanisms of fibrosis: therapeutic translation for fibrotic disease. *Nat Med* 18, 1028-1040
103. Li XH, Xue WL, Wang MJ et al (2017) H<sub>2</sub>S regulates endothelial nitric oxide synthase protein stability by promoting microRNA-455-3p expression. *Sci Rep* 7, 44807
104. Fouad AA, Hafez HM and Hamouda A (2020) Hydrogen sulfide modulates IL-6/STAT3 pathway and inhibits oxidative stress, inflammation, and apoptosis in rat model of methotrexate hepatotoxicity. *Hum Exp Toxicol* 39, 77-85
105. Zeng J, Lin X, Fan H and Li C (2013) Hydrogen sulfide attenuates the inflammatory response in a mouse burn injury model. *Mol Med Rep* 8, 1204-1208
106. Mao YQ and Fan XM (2015) Autophagy: A new therapeutic target for liver fibrosis. *World J Hepatol* 7, 1982-1986
107. Singh KK, Lovren F, Pan Y et al (2015) The essential autophagy gene ATG7 modulates

organ fibrosis via regulation of endothelial-to-mesenchymal transition. J Biol Chem 290, 2547-2559

108. Lucantoni F, Martinez-Cerezuela A, Gruevska A et al (2021) Understanding the implication of autophagy in the activation of hepatic stellate cells in liver fibrosis: are we there yet? J Pathol 254, 216-228
109. Lv S, Liu H and Wang H (2021) Exogenous Hydrogen Sulfide Plays an Important Role by Regulating Autophagy in Diabetic-Related Diseases. Int J Mol Sci 22

## FIGURE LEGENDS

**Figure 1. Various cellular functions of H<sub>2</sub>S in the liver.** Three major enzymes responsible for H<sub>2</sub>S production are CBS, CSE, and MPST. L-cysteine is the major substrate for H<sub>2</sub>S production. H<sub>2</sub>S-mediated signaling varieties from protein modification by sulfidation to affecting a broad range of physiological processes, including regulation of mitochondrial biogenesis, glucose metabolism, oxidative stress, inflammation, fatty acid oxidation and crosstalk with other signaling molecules. CBS: cystathionine  $\beta$ -synthase; CSE: cystathionine  $\gamma$ -lyase; MPST: 3-mercaptopyruvate sulfur transferase.

**Figure 2. Endogenous and exogenous production of H<sub>2</sub>S in the liver and its effects on liver fibrosis.** H<sub>2</sub>S plays a complex role in the development of fibrosis. Besides as a reducer to directly scavenge reactive oxygen species, exogenous (NAHS, GYY4137) or endogenous H<sub>2</sub>S utilizes its inhibitory effect on fibrosis by anti-inflammation and suppression of fibroblasts activation. Many signaling pathways, such as TNF- $\alpha$ , NF- $\kappa$ B, MAPKs, NRF2, SIRT1, SIRT3, GSH, TGF- $\beta$ 1/SMAD, PI3K, AKT, and autophagy are involved in the process of antifibrosis of H<sub>2</sub>S. TNF- $\alpha$ : tumor necrosis factor-alpha; NF- $\kappa$ B: nuclear factor-kappa B; mitogen-activated protein kinase: MAPK; NRF2: nuclear factor erythroid 2-related factor 2; SIRT1: sirtuin 1; SIRT3: sirtuin 3; GSH: glutathione; TGF- $\beta$ 1: transforming growth factor beta 1; SMAD: suppressor of mothers against decapentaplegic; PI3K: phosphoinositide 3-kinase.

Fig.1

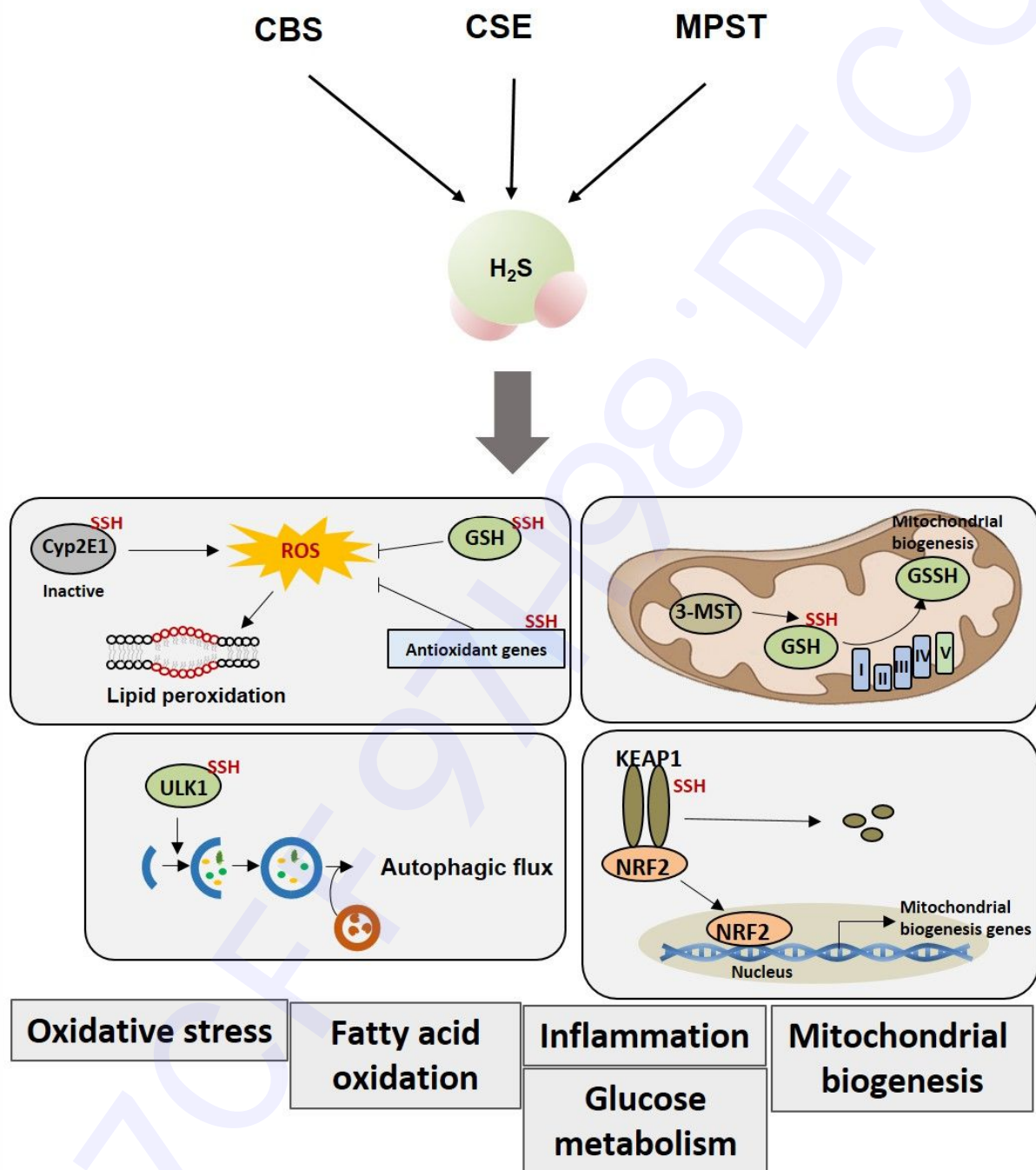




Fig.2

