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#### **Mini Review**

The soma-germline communication: Implications for somatic and reproductive aging

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#### **ABSTRACT**

Aging is characterized by a functional decline in most physiological processes, including alterations in cellular metabolism and defense mechanisms. Increasing evidence suggests that caloric restriction extends longevity and retards age-related diseases at least in part by reducing metabolic rate and oxidative stress in a variety of species, including yeast, worms, flies, and mice. Moreover, recent studies in invertebrates – worms and flies, highlight the intricate interrelation between reproductive longevity and somatic aging (known as disposable soma theory of aging), which appears to be conserved in vertebrates. This review is specifically focused on how the reproductive system modulates somatic aging and vice versa in genetic model systems. Since many signaling pathways governing the aging process are evolutionarily conserved, similar mechanisms may be involved in controlling soma and reproductive aging in vertebrates.

#### **INTRODUCTION**

The central role of oxidative damage, telomere attrition, mitochondrial genome damage, and epigenetic alterations in aging is well documented (1). The study of these different causes is best done through the use of model organisms as they are readily amenable to both genetic manipulation and molecular analysis. Two simple model organisms where aging is well studied are yeast (*Saccharomyces cerevisiae*) and worms (*Caenorhabditis elegans*) (Fig. 1). The aging of yeast can be monitored in two distinct ways: replicative lifespan and chronological lifespan (2). Replicative lifespan in yeast is determined by the number of divisions that each mother cell can undergo, whereas chronological yeast aging is measured using stationary phase cultures (2). In *C. elegans*, lifespan is measured by the amount of average days that the animals live and, as adult worms do not undergo further somatic cell division, aging is post-mitotic. Both of these model organisms have remarkably similar viability curves and mounting evidence indicates common characteristics of aging with mammals (3).

Genetic studies in model organisms have shown that aging is regulated by a specific cluster of genes and have allowed for the analysis of the different pathways involved in physiology, signal transduction, and gene regulation (4). Of note, energy depletion (and calorie restriction) in yeast through glucose deprivation was found to extend the lifespan of mother cells (5). This is due to silencing of DNA via deacetylation of histones by silent information regulator (Sir) proteins (4) (Fig. 1). These proteins are downstream of a signaling pathway involving Sum1p that detects cellular glucose and responds by repressing key genes that promote aging (4, 6). Homologs of the Sir proteins (Sir-2.1) have also been studied in *C. elegans* where the upregulation of a Sir homolog was able to extend lifespan (7, 8). Like *C. elegans, Drosophila melanogaster* (*D. melanogaster*) also exhibits biological properties suitable for aging studies such as high fecundity, short generation time, ease of cultivation,

and availability of sophisticated methods for genetic studies (9) (Fig. 1). These attributes also compound with the fact that the role of the insulin/IGF-1 (insulin-like growth factor-1) pathway in aging is evolutionarily conserved (4). Along those lines, findings by Tatar *et al.* suggest that longevity in *D. melanogaster* is controlled by the ratio of consumed protein relative to carbohydrates (10). This nutrient sensing is thought to occur through insulin-like peptides that regulate longevity. Lifespan extension parallels between *C. elegans* and *D. melanogaster* have also been found between *C. elegans* insulin/IGF-1 receptor gene, *daf-2 (dauer formation-2)*, and *D. melanogaster* insulin/IGF-1 receptor, INR (insulin-like receptor) (11) (Fig. 1). In fact, blocking *daf-2* expression inhibits PI3K/Akt (Phosphatidylinositol 3-kinase/protein kinase B) pathway, and promotes longevity (12). Very recently, the essential role of INR was highlighted by Ma *et al.* who showed that probiotics significantly enhance the lifespan of wild-type *D. melanogaster* but does the opposite in *InR*<sup>[E19]</sup> *Drosophila* (13).

The mouse model is also an attractive choice for aging studies. Mice have a relatively shorter lifespan and a wide array of anatomical and physiological characteristics that are shared with humans (14). In mice, calorie restriction similarly delays aging at least in part through SIRT1-dependent PGC- $1\alpha$  (peroxisome proliferator-activated receptor gamma coactivator  $1\alpha$ ) activation (15) and SIRT1-mediated antioxidant response in mice (16) (Fig. 1). On the contrary, upregulation of insulin/IGF-1 accelerates aging and predisposes to agerelated diseases (17) (Fig. 1). Mice have long telomeres and higher telomerase activity in many organs, which limits the usefulness of comparative studies with humans (14). Using telomerase-deficient mice, it has been demonstrated that telomere damage promotes age-associated decline in organ function and increased disease risk (18). Moreover, telomere dysfunction accelerates aging in mice and humans, as evidenced by the delay in aging upon experimental stimulation of telomerase in mice (1). However, excessive telomerase activity is a critical step for the development of human cancers. Notably, calorie restriction also

attenuates aging-associated shrinkage of telomeres in mouse tissue and reduces the incidence of tumors in mice that overexpress telomerase (19). Therefore, molecular and cellular mechanisms controlling aging and aging-associated pathology are likely conserved across species.

#### 1. Soma Theory of Aging

A key principle that describes the trade-off between immortal germline and the disposable soma is Thomas Kirkwood's disposable soma theory of aging. This theory proposes that germline immortality comes at the expense of somatic aging (Fig. 2A and 2B). Also, the removal of germline delays aging and improves somatic recovery (20, 21) (Fig. 2C). Thus, the soma is disposable and the resources are shifted to be used on reproduction, and the germline is labeled as expensive (22). While there has been work supporting this theory, another view challenged the disposable soma theory. Rather than the soma being disregarded so that more resources are focused on germline maintenance, these two components work together through common signaling pathways (22).

The disposable soma theory has been supported through a countless number of experiments using model organisms such as worms, flies, and zebrafish. In regard to *C. elegans*, studies using Catechin and tannic acid demonstrated a decrease in body size in exchange for longevity which argues for the disposable soma theory (23, 24). *D. melanogaster* is another model organism that demonstrates this trade-off, with juvenile hormone (JH) as a regulator (25), which will be discussed further in the section on the somatic effect on reproductive aging. In addition, late-reproducing fly strain had a significantly longer lifespan than early-reproducing strain (26). In male zebrafish, the experimental removal of the germline improves somatic recovery in response to stress (21) (Fig. 2C).

As for humans, Min *et al.* reported the effects of castration and Korean eunuchs on longevity. It was found that the average lifespan of 70 years observed in Korean eunuchs was significantly lowered in non-castrated men who had an average of 50-56 years (27). It was then proposed that having male sex hormones reduced longevity; again supporting the disposable soma theory (27).

#### 2. Reproductive Effect on Somatic Aging

To date, significant progress has been made in understanding genetic and environmental factors of aging. Likewise, the interest in the molecular mechanisms through which the reproductive system modulates somatic aging and vice versa, have recently emerged in multiple model systems. C. elegans have a two-part reproductive system which entails that lifespan is dictated by both germ cells and somatic reproductive tissues (28). C. elegans germ cells are totipotent and constitute the only immortal lineage capable of generating offspring. Somatic cells, however, differentiate into specialized, mortal cell types (29). In order for the germline stem cells (GSC) to achieve immortality, they must exhibit exquisite maintenance and repair machinery (30). In a study by Lee et al., it was found that the germline promotes longevity at 15°C. Low temperatures are speculated to decrease the damage and deterioration of distinct tissues triggered by cellular metabolism leading to an overall healthier soma (31). Along with this decrease in cellular damage, it is also suggested that the germ cells activate distinct signals to differentially modulate somatic tissues depending on physiological and environmental conditions (18). Uncovering these mechanisms of somatic tissues adopting a more germ cell-like character as well as taking in signals from GSCs may provide a means of dissecting the dichotomy between the instability of somatic cells and the immortality of GSCs (32). Synergistic lifespan extension is observed in a double mutant of DAF-2 (an ortholog of human insulin receptor) and RSKS-1 (an

ortholog of human ribosomal protein S6 kinase), which produced nearly a five-fold increase in longevity. In fact, inhibition of RSKS-1 activates DAF-16 (an ortholog of human FOXO (forkhead box O1)) in the intestine; a master transcriptional regulator involved in stress response against aging. The positive feedback of DAF-16 was mediated through AMPactivated kinase (AMPK) in daf-2 and rsks-1 mutants (33). In a study by Yamawaki et al. (28), the removal of germ cells resulted in nuclear accumulation of the transcription factor DAF-16. When germ cells are not removed, DAF-2 has the ability to inhibit DAF-16 by phosphorylation and thus prevent it from nuclear accumulation leading to reduced lifespan. Nuclear accumulation of DAF-16 has also been shown to be stimulated not only by removal of germ cells but also through the use of hydrostatic pressure. Furthermore, Watanabe et al., found cyclic pressure treatment (of 1MPa once a day for 5 minutes from L1 larvae until death) to significantly increase C. elegans lifespan; an event similarly associated with nuclear translocation of DAF-16 (34). Along those lines, it has very recently been demonstrated that DAF-16 does similarly mediate chemically-induced lifespan extension as caused by the terpenoid 3,3-dimethylallyl alcohol (Prenol) (35). This highlights the role of DAF-16 in stress response, adaptive behavior, and regulation of longevity.

Likewise, the effect of germline ablation on longevity was also examined in D. *melanogaster*. The idea of this was based on the study of sterile grandchildless mutants of D. *subobscura* and the finding that such mutants had an extended lifespan (36). GSC- loss is caused by overexpressed  $bam^+$  (37). The ablation of the germline resulted in an increase in lifespan by 31.3% to 50% in females and 21% to 27.8% in males (37). Also observed was that germ cell overproliferation shortens the life of the organism (37). Aging delay and increased lifespan was due to the elimination of metabolic demands required to produce gametes, as a result of the removal of the germline (37).

Other studies have focused on intermediate metabolites of the pyrimidine metabolism pathway to infer how it regulates reproductive signals involved in lifespan of *C. elegans*. Extension of lifespan through this pathway, with mediators such as thymine, functions to inhibit reproductive signals and subsequently induce DAF-12 (nuclear receptor of dafachronic acid hormone), NHR (nuclear hormone receptor family), and DAF-16 (38). In another study by Wan *et al.*, it was shown that thymine treatment affected fat metabolism, which may influence aging and longevity. A study by Ratnappan *et al.* suggests that NHR-49 may facilitate the adaptation to loss of reproductive potential through synchronized enhancement of fatty acid oxidation and desaturation (39). This study also proposes that the conversion of a saturated fatty acid-rich lipid profile to a monounsaturated fatty acid- and polyunsaturated fatty acid-enriched one may lead to enhanced somatic maintenance and lipid signaling, eventually increasing longevity.

Along with lipid metabolism, autophagy is another important lifespan extension mechanism in multiple animals (Fig. 1). Specifically, in *C. elegans*, germline-loss has also been observed to trigger Target of Rapamycin (TOR) downregulation which stimulates PHA-4 (an ortholog of human Forkhead box A) and autophagy (40). According to a study by Lapierre and Hansen, autophagy was required for the increased lipase activity in animals without germline, which supports the existence of a functional link between autophagy and lipid breakdown (41). As seen with the activation of DAF-16 in the intestine, Lapierre and Hansen found that autophagy is predominantly induced in hypodermal and intestinal cells which suggests these tissues are particularly important for longevity. Likewise, the longevity-promoting effects of the flavonoid 5'-Hydroxy-6, 7, 8, 3', 4'-pentamethoxyflavone (5-HPF) was through stimulation of autophagy as evident from the increase in transcriptional activity of *bec-1* (an ortholog of the human BECN1) and *lgg-1* (an ortholog of the human GABA type A receptor associated protein like 1) (42). In congruence with these findings, overexpression

of the autophagy receptor SQSTM1 (Sequestosome 1, also called p62) is sufficient to promote longevity in *C. elegans* and *D. melanogaster* (43). Notably, the conserved function of autophagy in longevity has been proved in multiple animals (Fig. 1).

In a study by Wei and Kenyon, the role of reactive oxygen species (ROS) and hydrogen sulfide in the longevity response to germline loss in *C. elegans* was examined. With regard to ROS activation, it is thought that germline removal is a signal that germline maturation is incomplete and in response attempts to generate more energy for germline biosynthesis by increasing electron transport rates (44). The hydrogen sulfide pathway was also upregulated by germline-loss and when this pathway was blocked a decrease in longevity in worms without germline was observed (44).

Prostaglandins have also been shown to be involved in longevity and lifespan determination. It was found that a relationship existed between lower, optimal body temperature, germline, and lifespan in *C. elegans*, as discussed in the study published by Lee *et al.* (31). The GSCs respond to the decrease in body temperature by releasing prostaglandin E2 (PGE2), which in turn instigates intestinal expression of *cbs-1* (cystathionine beta-synthase-1) (31). The intestine then produces more hydrogen sulfide, which may explain the delay in somatic aging in this organism (31). Although colder temperatures are preferred, overexpression of *cbs-1* also extends lifespan at warmer temperatures (31).

#### 3. Somatic Effect on Reproductive Aging

In wild-type *C. elegans*, reproduction and aging are inversely related (8). Of note, inhibition of insulin/IGF-1 and TGF- $\beta$  (transforming growth factor- $\beta$ ) Sma/Mab pathways has been shown to slow down reproductive aging. The TGF- $\beta$  Sma/Mab pathway functions in the hypodermis to regulate body size autonomously (45). Intriguingly, Luo *et al.* have

reported that reduction of both pathways attenuates reproductive aging by maintaining oocyte and germline quality non-cell autonomously (46).

DAF-12 is a receptor in *C. elegans* that impacts lifespan by determining whether induction of reproductive growth, its arrest, or dauer diapause, should occur (47). The arrested state of dauer diapause is chosen in poor growth conditions. 3-kelo bile acid-like steroids, which are comprised of Δ4-dafachronic acid and Δ7-dafachronic acids, act on DAF-12 receptor (48). One of influencers for a decision making between dauer diapause and reproductive growth is DAF-9 (an ortholog of human cytochrome P450 family) (48). DAF-9/P450 produces dafachronic acid ligands that activate the DAF-12 nuclear receptor, directing reproductive growth (48-50). Notably, DAF-9 and DAF-12 act through DAF-16 to extend the lifespan of germline-loss worms (48-50).

Studies have shown that there is an inverse relationship between aging and GSC proliferation. As *D. melanogaster* ages, significant reduction in the rate of GSC proliferation is observed (51). It appears that there are parallel processes that contribute to the maintenance of gametocytes and proliferation of the germline in *C. elegans* and *D. melanogaster*. Similar to *C. elegans*, *D. melanogaster* also expresses *dpp* which encodes a protein of the TGF-β family that may be involved in reproductive aging if alterations occur such as changes to the GSC niche (52). However, *D. melanogaster* TGF-β is released from the stem cell niche, which is somatic. The expression of this gene in niche cells starting at a young age results in enhanced egg production at later stages of life; it does so by inhibiting differentiation of cystoblasts in germlines (53). Thus, deterioration of reproduction is also influenced by somatic tissues (51). *D. melanogaster* also expresses insulin-like peptide, which is derived from neurosecretory cells located in the brain. This protein normally upregulates GSC proliferation following high-protein intake (54). It has been found that removal of Insulin-like peptides in the brains of young adult female *D. melanogaster* slows down GSC proliferation

(55). Additionally, the inhibition of the gonad's insulin receptors delays oogenesis (51). Notably, TGF-β signaling has been implicated in multiple aspects of mammalian reproductive aging (56). This suggests that TGF-β Sma/Mab and Insulin/IGF-1 are ancestral somatic signaling pathways that govern, at least in part, reproductive aging.

Several studies have been done focusing on JH and its effects in different invertebrates. JH is involved in the metamorphosis of this organism by stimulating differentiation and inhibiting morphogenesis, or the formation of tissues (57). The researchers proposed that JH was a mediator for reproductive and longevity trade-off (25). There is also an inverse relationship on reproduction, which is caused by JH, and survival (58). This signaling pathway occurs downstream of the Insulin/IGF-1 pathway (25). JH analog (JHa) methoprene was used for selection, to mimic the effects of JH-deficiency in flies. As expected, JHa treatment increased mortality in the unselected, susceptible controls in exchange for early fertility (25). Upon successful selection, flies gained resistance to JHa, extending their lifespan by an average of 3.8 days (25). This finding was in agreement with the longer lifespan in flies without JH (25).

It is established that hormones are essential in regulating body processes, and germline aging is no exception. The anterior pituitary gland is a major organ of the endocrine system that secretes gonadotropins, such as luteinizing hormone (LH). LH plays an important role in reproductive aging in mammals (59). A preovulatory LH surge causes oocytes to be arrested during prophase I of meiosis in females. Likewise, LH also increases the survival rate of male germ cells (60). In a study by Kawamura *et al.*, a regulatory mechanism controlled by the anterior pituitary gland of *Rattus norvegicus* was demonstrated. In particular, activation of the Gi pathway caused significant reduction in cAMP (cyclic adenosine monophosphate) levels (60). Additionally, administration of Leydig insulin-like 3 (INSL3) suppressed programmed cell death in male germ cells and prevented the arrest of

oocytes at prophase I (60). Furthermore, glial cell line-derived neurotrophic factor (GDNF) is released from somatic Sertoli cells of *Mus musculus* to influence self-renewal, differentiation, and maintenance of GSCs, as demonstrated by premature depletion of GSCs in *gdnf* <sup>+/-</sup> mice (61). Other signals originating from the Sertoli cells are bone morphogenetic protein 4 (BMP4) and stem cell factor (SCF) which contribute to germ cell maintenance and differentiation, respectively (62).

#### 4. Conclusion

Mounting evidence suggests that somatic tissues play a critical role in reproductive aging and vice versa. Using animal models such as *C. elegans*, *D. melanogaster*, *R. norvegicus*, and *M. musculus*, the interaction between somatic tissue and germline can be extrapolated to other organisms, including humans. It was also widely accepted that organisms allocate limited resources (e.g., energy) to produce either more offspring (reproduction) or to live longer (soma tissue maintenance). The disposable soma theory of aging proposed by Kirkwood states that it is evolutionarily beneficial to sacrifice the soma tissue maintenance (longevity) for the sake of the reproduction. Although many animal models including humans appear to support this theory, it are still controversial (63) - Ermolaeva *et al.* found that a systemic response to DNA damage in germ cells protects somatic tissues against multiple stresses (64). Therefore, we suggest that the soma-germline communication on aging may be a unidirectional and a bidirectional, depending on external and intrinsic conditions.

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## **CONFLICTS OF INTEREST**

The authors declare no conflict of interest.

#### FIGURE LEGENDS

**Figure 1. A conserved pathway for aging in several species.** SIR2 homologs (yeast Sir2, worm sir-2.1, fly dSir2, and mouse SIRT1) activated by calorie restriction inhibit aging through FOXO homologs and autophagy. In contrast, Insulin/IGF-1 pathway promotes aging by inhibiting FOXO and autophagy but enhancing oxidative stress.

Figure 2. Communication between reproduction and soma. (A) Disposable soma theory of aging. An organism's lifespan (or aging) may be determined by an evolutionary trade-off between reproduction and tissue maintenance. (B) A greater consumption of a limited amount of resources (e.g., energy) in reproduction may result in reduced investment in tissue maintenance leading somatic aging. (C) Removal of reproductive tissue or reducing its activity delays aging processes in multiple organisms (e.g., Korean eunuchs), including humans.

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#### Yeast Worms Flies Mice Calorie Restriction (CR) DAF-2/IGF-1 CR Insulin/IGF-1 pathway Insulin/IGF-1 pathway CR CR Sir-2.1 AKT dSir2 SIRT1 Sir2 DAF-16 /FOXO Auto-phagy FOXOs -> Auto-AutodFOXO → Auto-phagy ROS phagy phagy phagy Aging **Aging Aging Aging**

Fig. 1. Figure 1

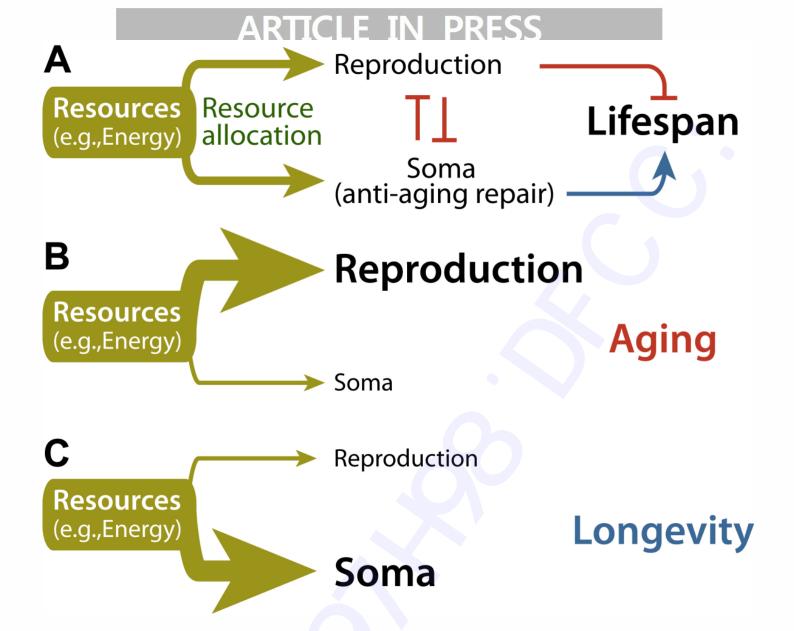


Fig. 2. Figure 2